

# **Spatial Rule-based Modeling of Cellular Biochemistry with MCell/ BioNetGen/CellBlender**

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GLBIO 2013 Tutorial, May 12

Goto <http://bionetgen.org/index.php/GLBIO2013> for slides and other materials

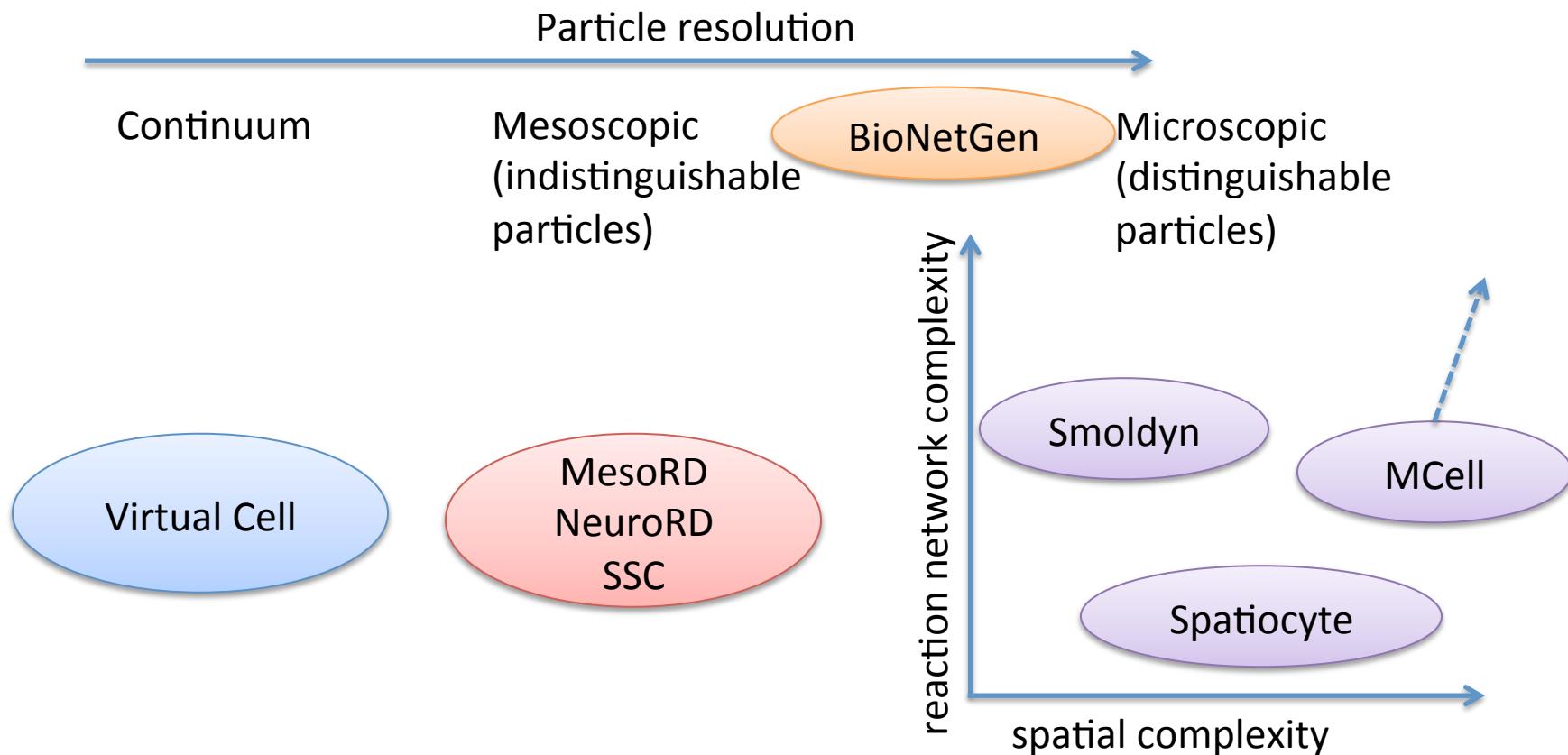
# Overview

- Multiscale Challenge (Dittrich)
- Intro to National Center for Multiscale Modeling of Biological Systems (Faeder)
- BioNetGen Motivation and Intro (Faeder)
- BioNetGen/RuleBender Demo (Faeder)
- MCell Intro (Dittrich)
- Mcell/CellBlender Demo (Dittrich)
- Features in Progress and Q&A

**The Multiscale Computational Challenge**

Problem/ Method	Typical Application	Software Examples	Resolution (Scale)	Spatial Realism	Stochastic Realism	Time Step	Time Scale	Serial/ Parallel	Computer Time
Networks of Reactions/ Sets of ODEs	Metabolic or signaling pathways	Virtual Cell ECell, Gepasi XPPAUT	N/A (cell)	N/A	<none>	ms	ms - hrs	serial	minimal
Excitation/ Compartmental Circuit	Nerve signaling	NEURON GENESIS NEOSIM	μm - mm (cell - multicell)	low - medium	none	ms	ms - hrs	usually serial	usually low
Reaction Kinetics/ Stochastic	Gene regulation/transcription	BioSpice StochSim XPPAUT <b>MCell</b>	N/A (cell)	N/A	high	ms	ms - hrs	serial	low
3-D Reaction Diffusion/ Finite Element	Flow models, calcium dynamics	Virtual Cell FIDAP Kaskade	<μm (cell)	medium-high	<none>	μs - ms	μs - sec	either	low - high
3-D Reaction Diffusion/ Monte Carlo	Micro-physiological processes	<b>MCell</b> ChemCell SmolDyn	nm – mm (subcell - cell)	high	high	ps - ms	μs - sec	either	low - high
Macromolecular Machinery/GNM	Collective dynamics	GNM ANM	Å - 100 nm (complexes)	high	none	N/A	<ns – μs>	N/A (analytic)	minimal
Diffusion in Potential Field/Poisson-Nernst-Planck	Electrostatic interactions, ion channels	UHBD Delphi <b>CHARMM</b>	Å - nm (membrane proteins)	high (implicit solvent)	none	N/A	<ns – μs>	parallel	low - medium
Macromolecular Motions/Brownian Dynamics (BD)	Conformational dynamics (in flow fields)	<b>CHARMM</b> GROMOS UHBD	Å - nm (macro-molecules)	high (implicit solvent)	high	5 - 10 fs	<ns – μs>	parallel	medium - high
Molecular Structure/Molecular Dynamics (MD)	Conformational dynamics & free energies	<b>NAMD</b> <b>AMBER</b> <b>CHARMM</b> GROMOS	Å (macro-molecules)	exact (explicit solvent)	exact	1 - 2 fs	<ns – μs>	parallel	very high
Transition Dynamics/Quantum Chem. + Mol. Mech. (QC/MM)	Enzyme reactions (make/break bonds)	<b>pDynamo</b> (AMBER CHARMM)	Å (molecules)	exact (explicit solvent)	exact	1 - 2 fs	<ns – μs>	parallel	very high
Molecular Structure/Ab initio simulations	Solution of the Schrodinger equation	Gaussian	<Å (electrons - atoms)	exact	exact	N/A	N/A	parallel	highest

# Comparison with other cell simulation tools



Biomedical Technology Research Center (BTRC)

High Performance Computing  
for  
**Multiscale Modeling of Biological Systems**

*Overarching biological theme:*

- Spatial organization
- Temporal evolution

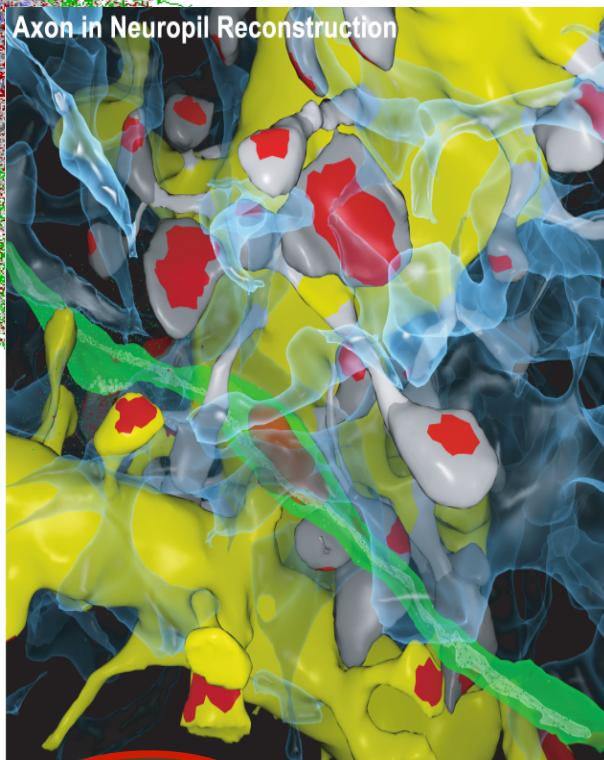
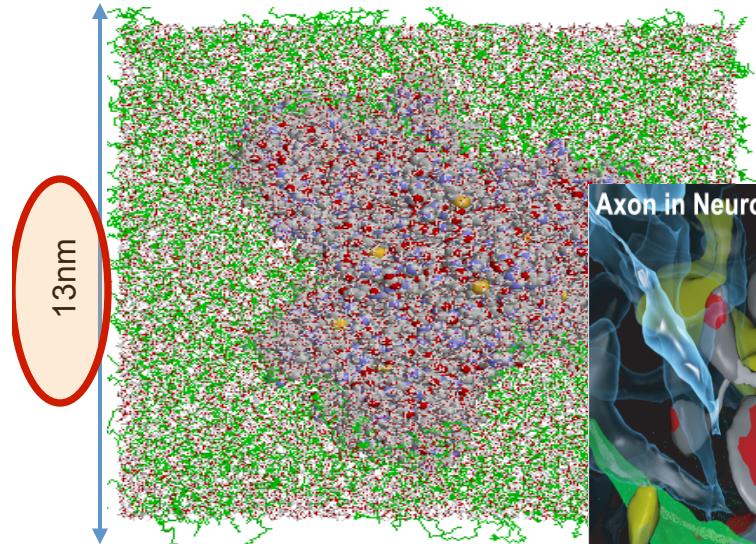
of (neuro)signaling systems/events



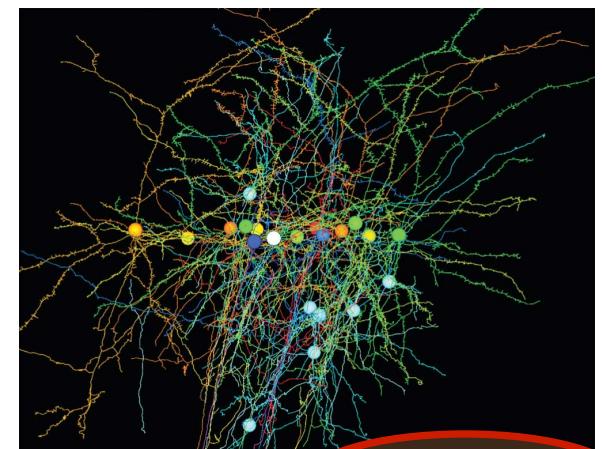
# From small molecules, to multimeric assemblies,

to cellular architecture,

to neural circuits

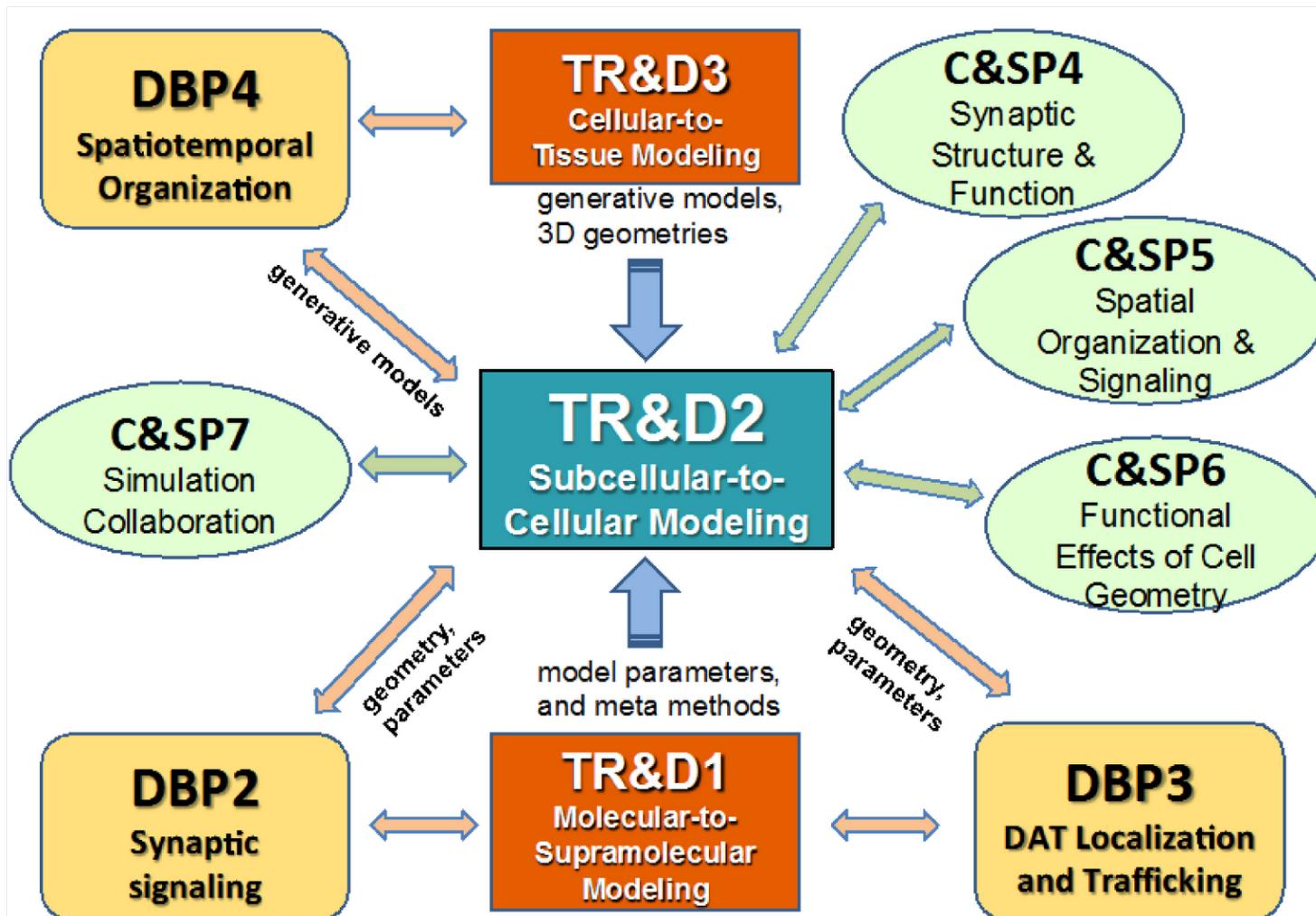


from  $6 \times 6 \times 5 \mu\text{m}^3$  sample of adult rat hippocampal stratum radiatum neuropil

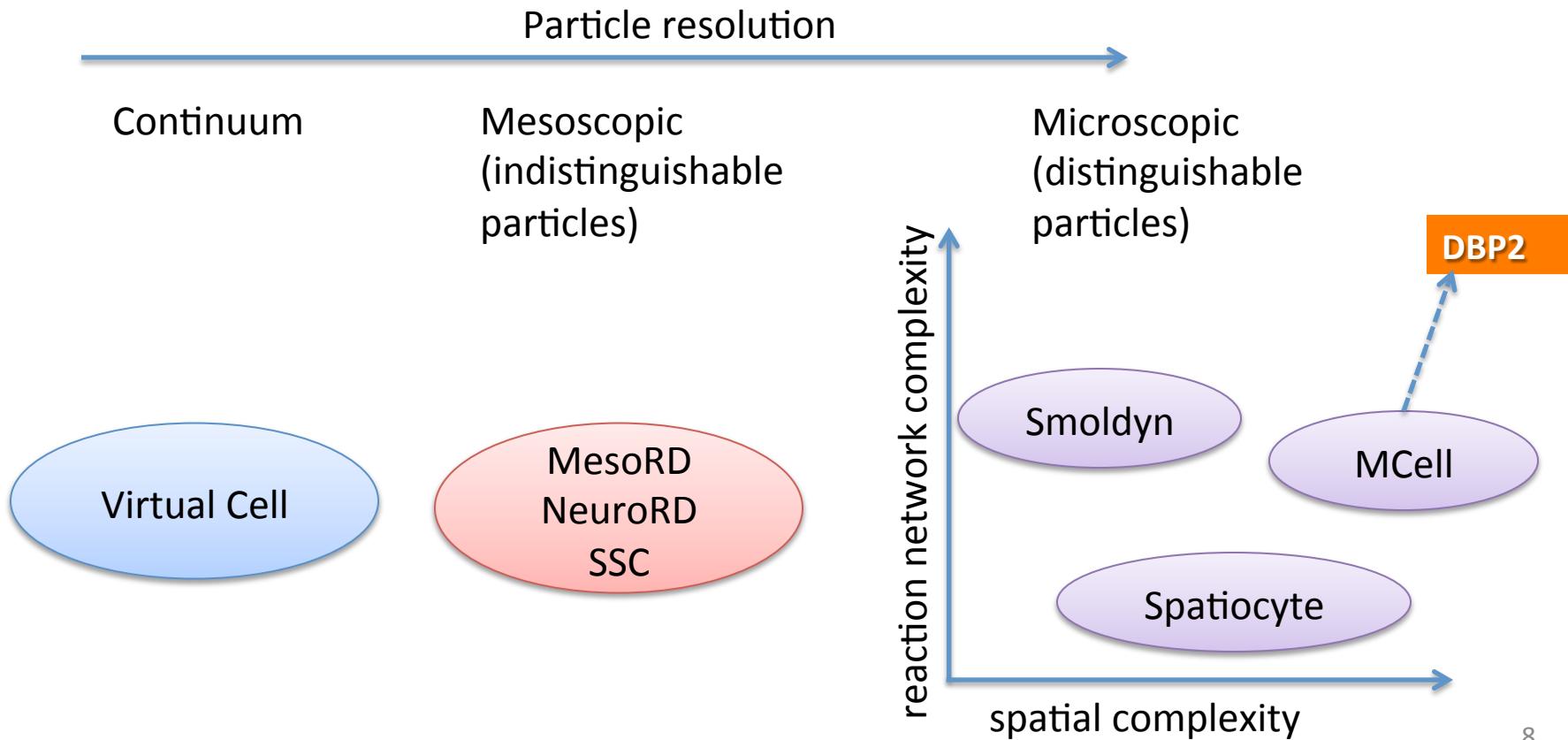


From SSEM images,  $400 \times 400 \times 50 \mu\text{m}^3$

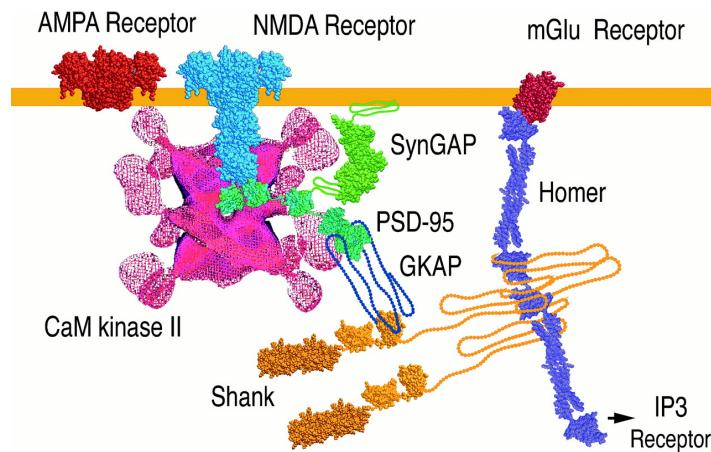
# Role of MCell in the BTRC



# Comparison of MCell with other tools for spatial modeling of biological systems



# Motivating example for Rule-Based Modeling



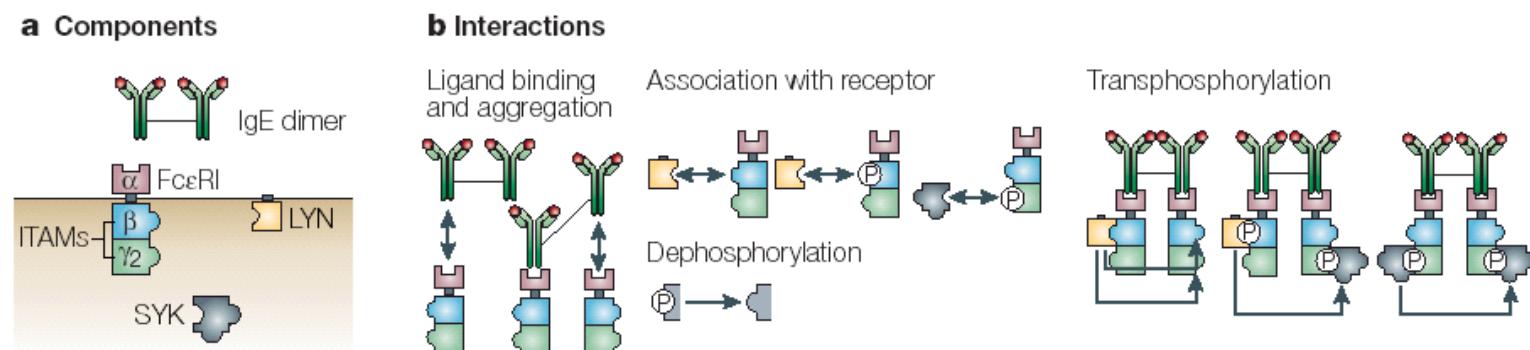
**Molecular machines in the PSD**

**Estimated number of states of  
CAMKII-CaM complexes:**

**40<sup>12</sup>**

# Standard modeling protocol

## 1. Identify components and interactions.

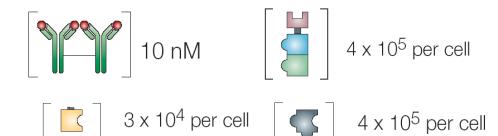


## 2. Write model reactions / equations

Reaction Network  $\longleftrightarrow \dot{\mathbf{x}} = \mathbf{S} \cdot \mathbf{v}(\mathbf{x})$

## 3. Determine **concentrations** and **rate constants**

## 4. Simulate and analyze the model



# Reactions to Differential Equations

Consider the reaction



The reaction rate is given by

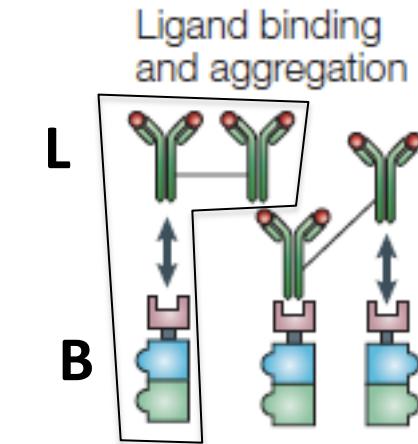
$$v_1 = k_1 R \cdot L$$

Rate of change of species concentrations (numbers) are

$$\frac{dR}{dt} = -v_1 + \dots$$

$$\frac{dL}{dt} = -v_1 + \dots$$

$$\frac{dRL}{dt} = +v_1 + \dots$$



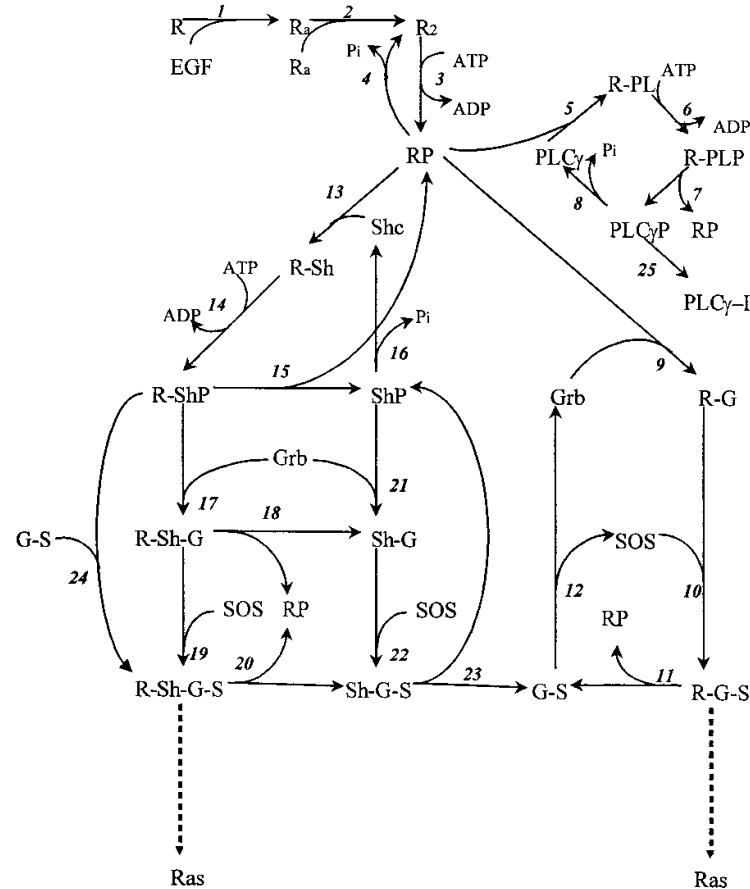
Here I have indicated that there may be additional terms from other reactions in the network. Reaction fluxes combine through *addition*.

# Reaction Network Models

Reaction Network Scheme



Mathematical Formulation



Rate Equations

$$\begin{aligned}
 & k_1 \cdot [R] \cdot [EGF] - k_{-1} \cdot [R_a] \\
 & k_2 \cdot [R_a] \cdot [R_a] - k_{-2} \cdot [R_2] \\
 & k_3 \cdot [R_2] - k_{-3} \cdot [RP] \\
 & V_4 \cdot [RP]/(K_4 + [RP]) \\
 & k_5 \cdot [RP] \cdot [PLC\gamma] - k_{-5} \cdot [R-PL] \\
 & k_6 \cdot [R-PL] - k_{-6} \cdot [R-PLP] \\
 & k_7 \cdot [R-PLP] - k_{-7} \cdot [RP] \cdot [PLC\gamma P] \\
 & V_8 \cdot [PLC\gamma P]/(K_8 + [PLC\gamma P]) \\
 & k_9 \cdot [RP] \cdot [Grb] - k_{-9} \cdot [R-G] \\
 & k_{10} \cdot [R-G] \cdot [SOS] - k_{-10} \cdot [R-G-S] \\
 & k_{11} \cdot [R-G-S] - k_{-11} \cdot [RP] \cdot [G-S] \\
 & k_{12} \cdot [G-S] - k_{-12} \cdot [Grb] \cdot [SOS] \\
 & k_{13} \cdot [RP] \cdot [Shc] - k_{-13} \cdot [R-Sh] \\
 & k_{14} \cdot [R-Sh] - k_{-14} \cdot [R-ShP] \\
 & k_{15} \cdot [R-ShP] - k_{-15} \cdot [ShP] \cdot [RP] \\
 & V_{16} \cdot [ShP]/(K_{16} + [ShP]) \\
 & k_{17} \cdot [R-ShP] \cdot [Grb] - k_{-17} \cdot [R-Sh-G] \\
 & k_{18} \cdot [R-Sh-G] - k_{-18} \cdot [RP] \cdot [Sh-G] \\
 & k_{19} \cdot [R-Sh-G] \cdot [SOS] - k_{-19} \cdot [R-Sh-GS] \\
 & k_{20} \cdot [R-Sh-G-S] - k_{-20} \cdot [Sh-G-S] \cdot [RP] \\
 & k_{21} \cdot [ShP] \cdot [Grb] - k_{-21} \cdot [Sh-G] \\
 & k_{22} \cdot [Sh-G] \cdot [SOS] - k_{-22} \cdot [Sh-G-S] \\
 & k_{23} \cdot [Sh-G-S] - k_{-23} \cdot [ShP] \cdot [G-S] \\
 & k_{24} \cdot [R-ShP] \cdot [G-S] - k_{-24} \cdot [R-Sh-G-S] \\
 & k_{25} \cdot [PLC\gamma P] - k_{-25} \cdot [PLC\gamma P-I]
 \end{aligned}$$

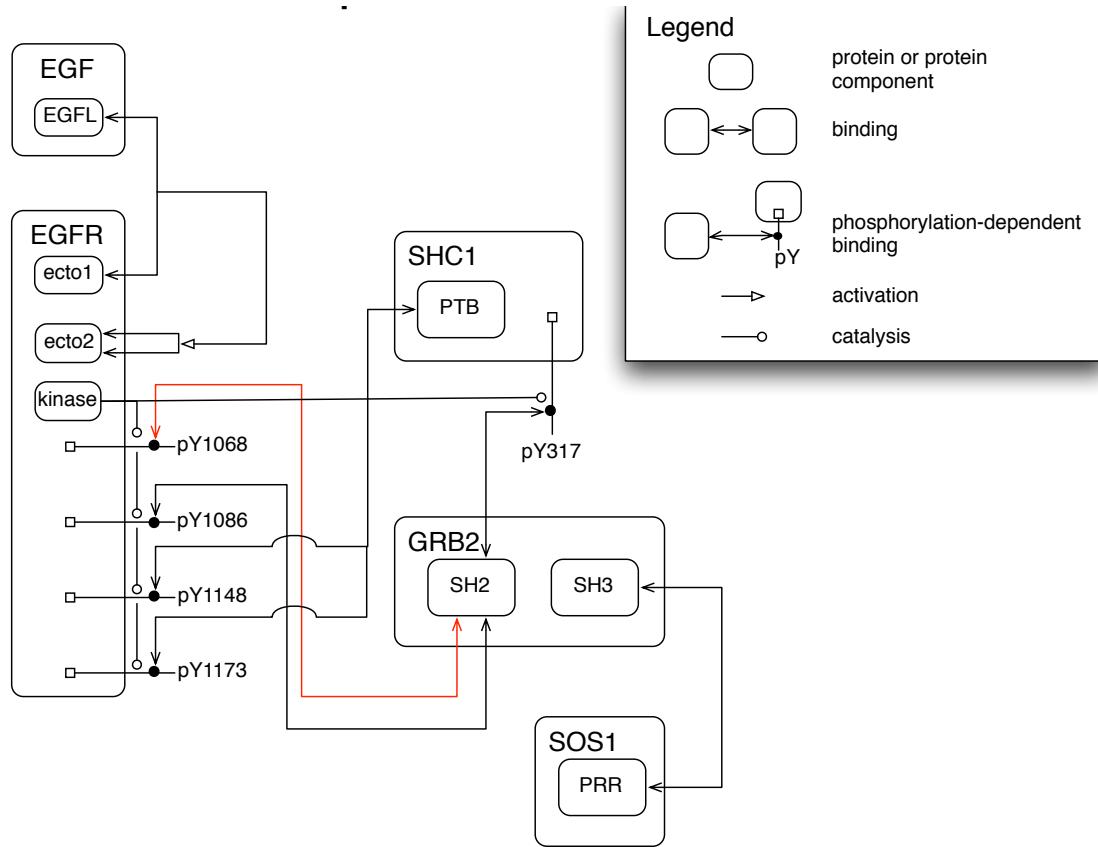
Differential Equations

$$\begin{aligned}
 d[EGF]/dt &= -v_1 \\
 d[R]/dt &= -v_1 \\
 d[R_a]/dt &= v_1 - 2v_2 \\
 d[R_2]/dt &= v_2 + v_4 - v_3 \\
 d[RP]/dt &= v_3 + v_7 + v_{11} + v_{15} + v_{18} + v_{20} - v_4 - v_5 - v_9 \\
 d[R-PL]/dt &= v_5 - v_6 \\
 d[R-PLP]/dt &= v_6 - v_7 \\
 d[R-Cl]/dt &= v_9 - v_{10} \\
 d[R-G-S]/dt &= v_{10} - v_{11} \\
 d[R-Sh]/dt &= v_{13} - v_{14} \\
 d[R-ShP]/dt &= v_{14} - v_{24} - v_{15} - v_{17} \\
 d[R-Sh-G]/dt &= v_{17} - v_{18} - v_{19} \\
 d[R-Sh-G-S]/dt &= v_{19} - v_{20} + v_{24} \\
 d[G-S]/dt &= v_{11} + v_{23} - v_{12} - v_{24} \\
 d[ShP]/dt &= v_{15} + v_{23} - v_{21} - v_{16} \\
 d[Sh-G]/dt &= v_{18} + v_{21} - v_{22} \\
 d[PLC\gamma P]/dt &= v_8 - v_5 \\
 d[PLC\gamma P-I]/dt &= v_{25} \\
 d[Grb]/dt &= v_{12} - v_9 - v_{17} - v_{21} \\
 d[Shc]/dt &= v_{16} - v_{13} \\
 d[SOS]/dt &= v_{12} - v_{10} - v_{19} - v_{22}
 \end{aligned}$$

22 species / 25 reactions

Kholodenko et al., *J. Biol. Chem.* (1999)

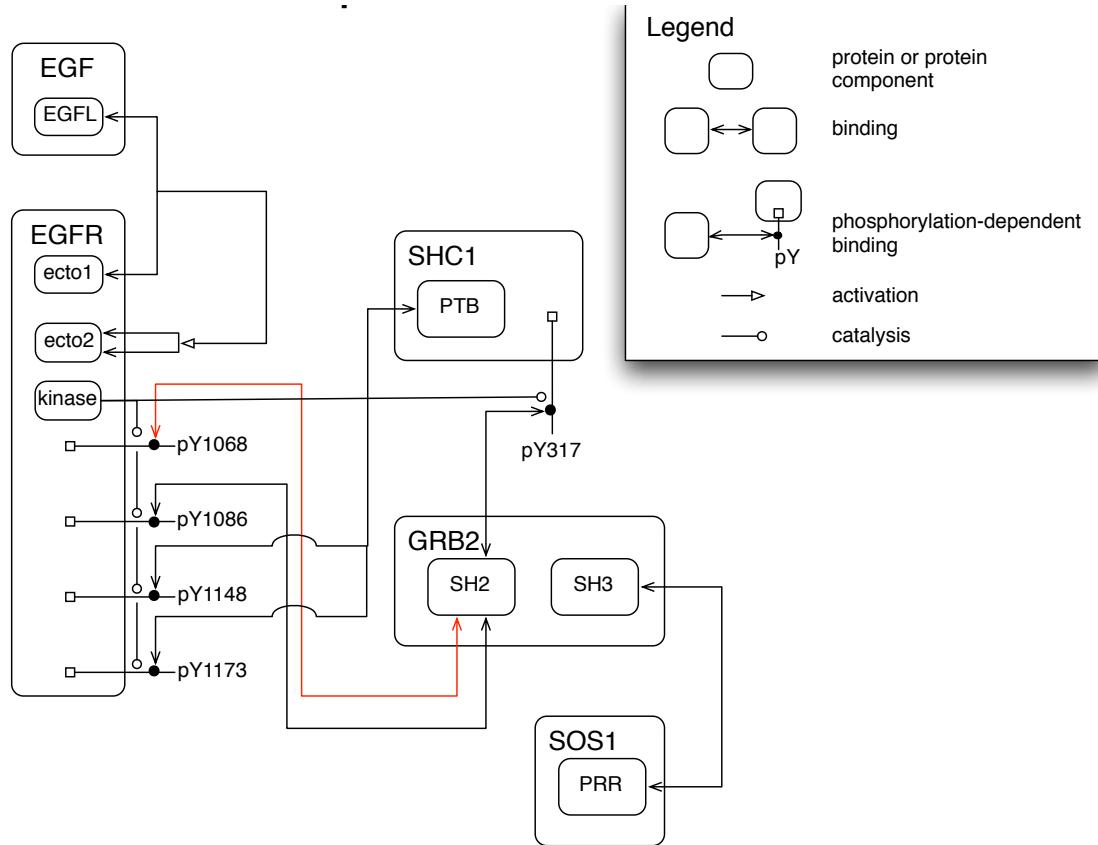
# Combinatorial complexity in a prototypical signaling module



5 proteins, 20 interactions → 170,000 species

... but only 20 (or so) rules.

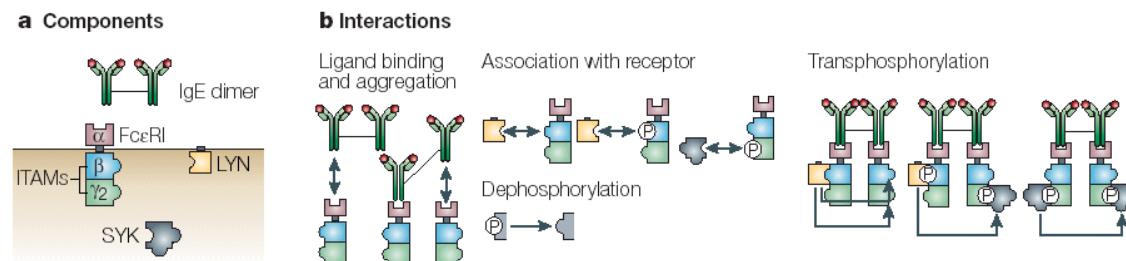
# Rules provide a scalable way to model molecular interactions



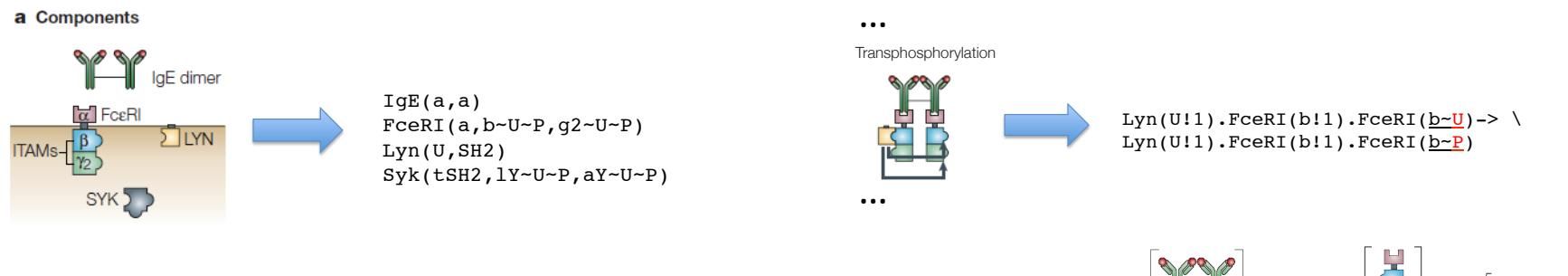
Rules ~ number of interactions << number of species

# Rule-Based Modeling protocol

## 1. Identify components and interactions.

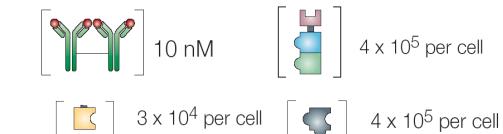


## 2. Translate into objects (molecules) and rules



## 3. Determine **concentrations** and **rate constants**

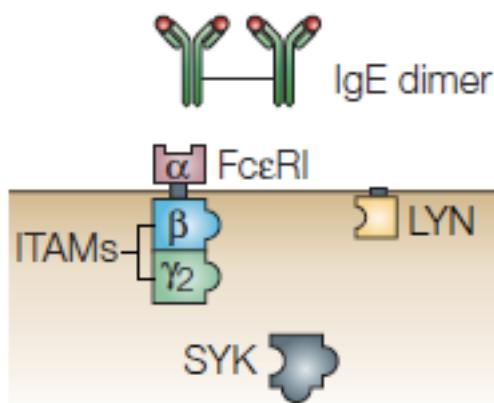
## 4. Simulate and analyze the model



# **SPECIFYING A RULE-BASED MODEL**

# Defining Molecules

Molecules are the basic objects in a BNG model



## BONETGEN Language

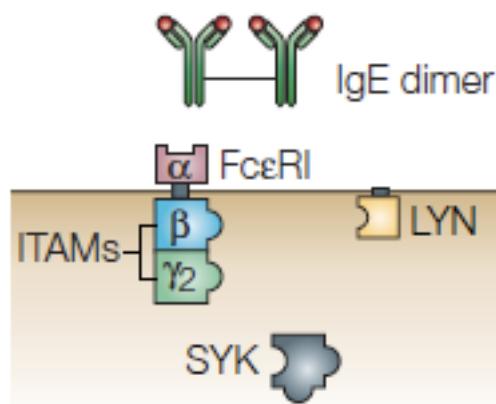
IgE(a,a)  
FcεRI(a,b~U~P,g2~U~P)  
Lyn(U,SH2)  
Syk(tSH2,1Y~U~P,aY~U~P)

**Components** represent molecule elements

- Domains
- Motifs
- Properties

# Defining Molecules

Molecules are the basic objects in a BNG model



## BIONETGEN Language

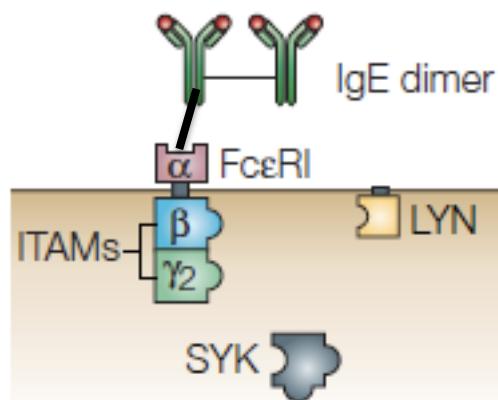
IgE(*a*,*a*)  
Fc $\epsilon$ RI(*a*,*b* $\sim$ U $\sim$ P,*g2* $\sim$ U $\sim$ P)  
Lyn(*U*,SH2)  
Syk(*t*SH2,*lY* $\sim$ U $\sim$ P,*aY* $\sim$ U $\sim$ P)

**Components** may have different **states** representing

- posttranslational modifications
- conformational state
- ...

# Binding

Molecules bind other molecules through components



## BIOGENE Language

$\text{IgE}(a, a!1).\text{Fc}\epsilon\text{RI}(a!1, b\sim U, g2\sim U)$

**Bonds** are formed by linking two components. The '.' indicates a set of molecules forming a complex.

$\text{Fc}\epsilon\text{RI}(a, b\sim U!1, g2\sim U).\text{Lyn}(U!1)$

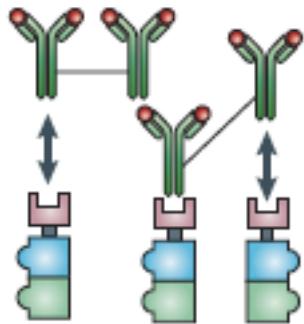
Components may have both states and bonds.

$\text{Lyn}(SH2!1, Cterm\sim P!1)$

Bonds may occur within a molecule.

# Defining Interaction Rules

Ligand binding  
and aggregation



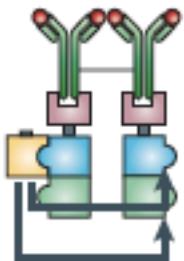
BIONETGEN Language

$\text{IgE}(a, \underline{a}) + \text{Fc}\epsilon\text{RI}(\underline{a}) \rightleftharpoons \text{IgE}(a, \underline{a!1}) \cdot \text{Fc}\epsilon\text{RI}(\underline{a!1})$

...

binding and dissociation

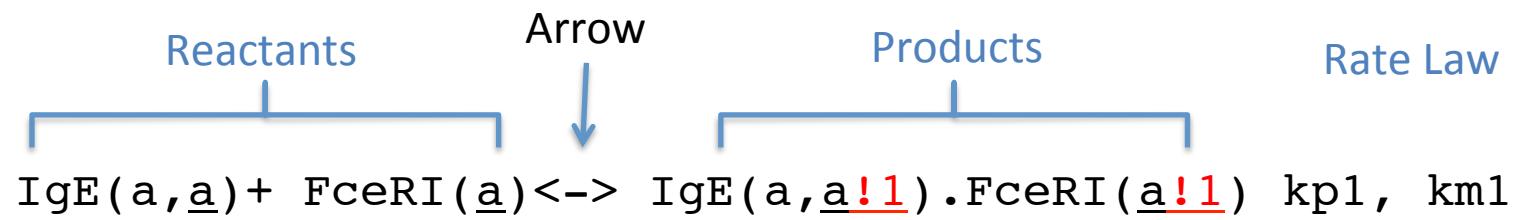
Transphosphorylation



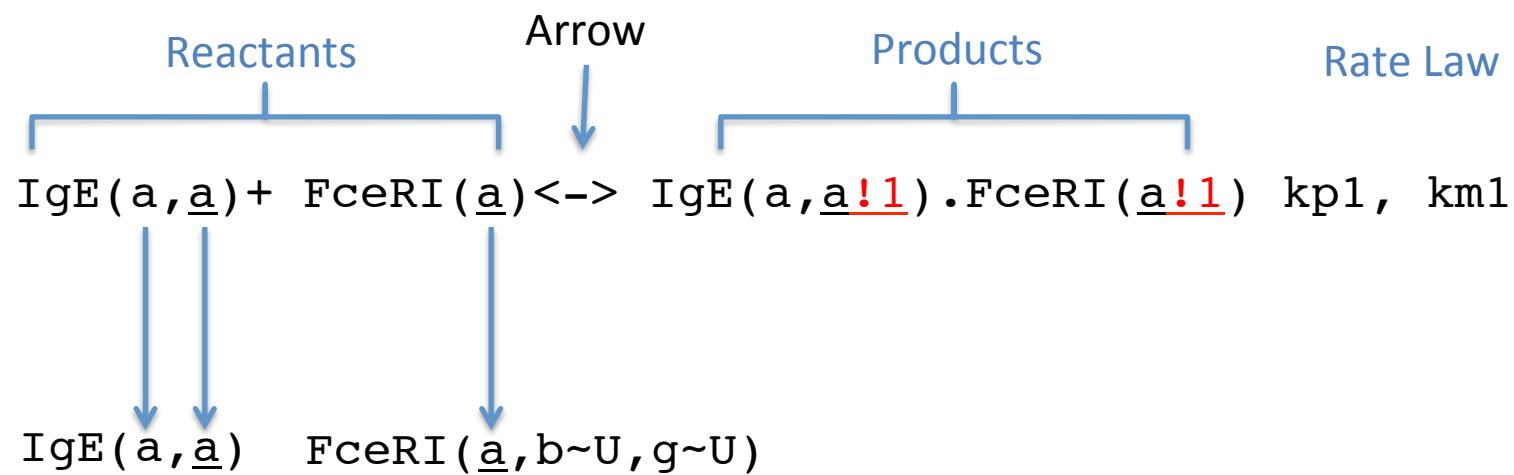
$\text{Lyn}(U!1) \cdot \text{Fc}\epsilon\text{RI}(b!1) \cdot \text{Fc}\epsilon\text{RI}(b\sim\underline{U}) \rightarrow \\ \text{Lyn}(U!1) \cdot \text{Fc}\epsilon\text{RI}(b!1) \cdot \text{Fc}\epsilon\text{RI}(b\sim\underline{P})$

component state change

# Parts of a rule

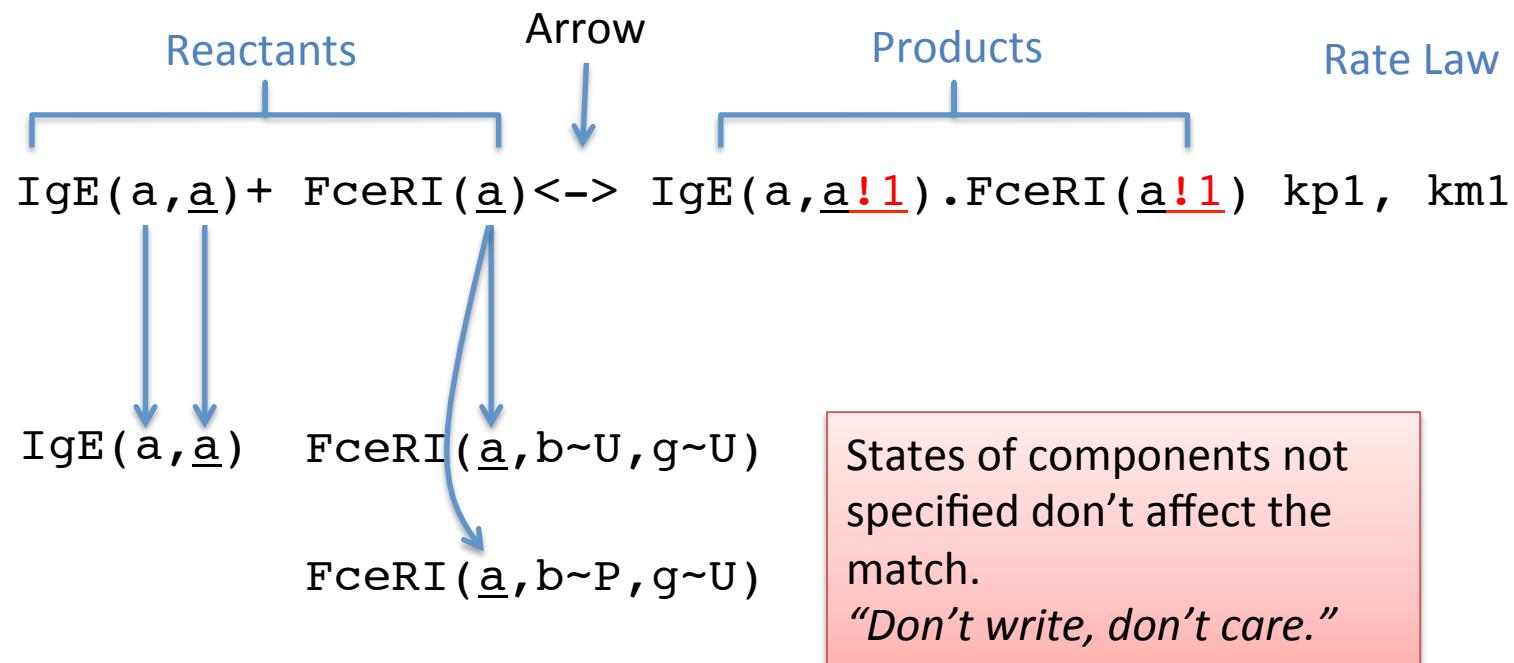


# Parts of a rule



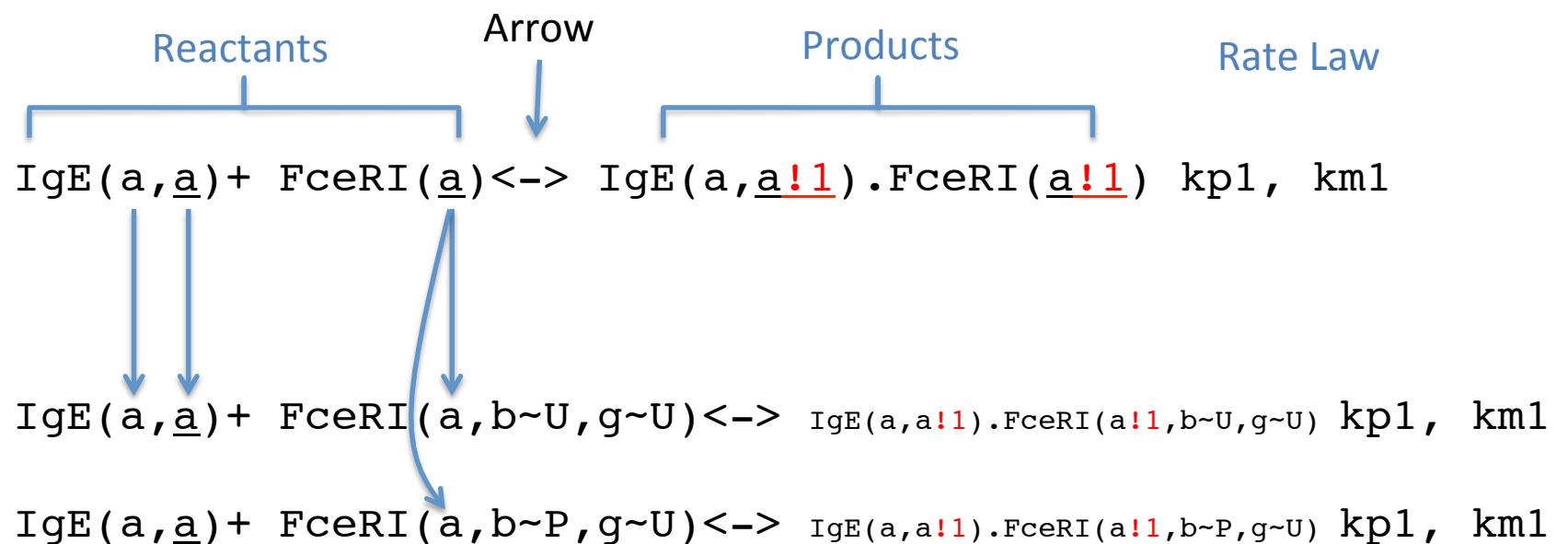
**Reactant patterns**  
select properties of  
each reactant  
molecule.

# Parts of a rule



**Reactant patterns**  
select properties of  
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# Parts of a rule

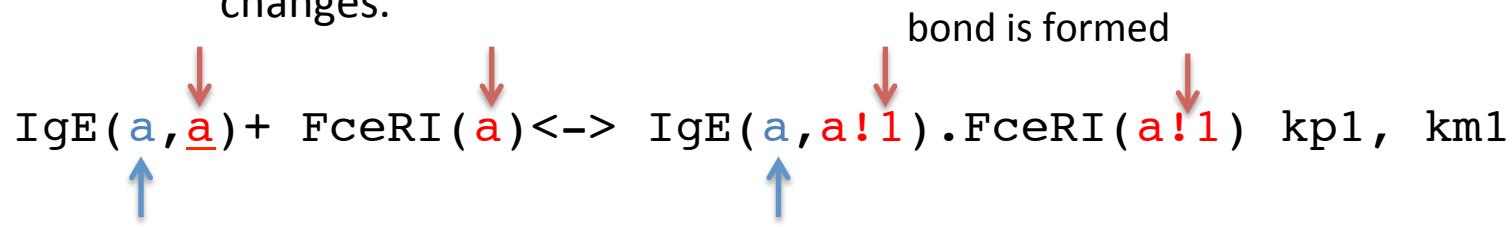


**Reactant patterns**  
select properties of  
each reactant  
molecule.

Because patterns can match  
many different species,  
each rule can generate  
many reactions.

# Center and context

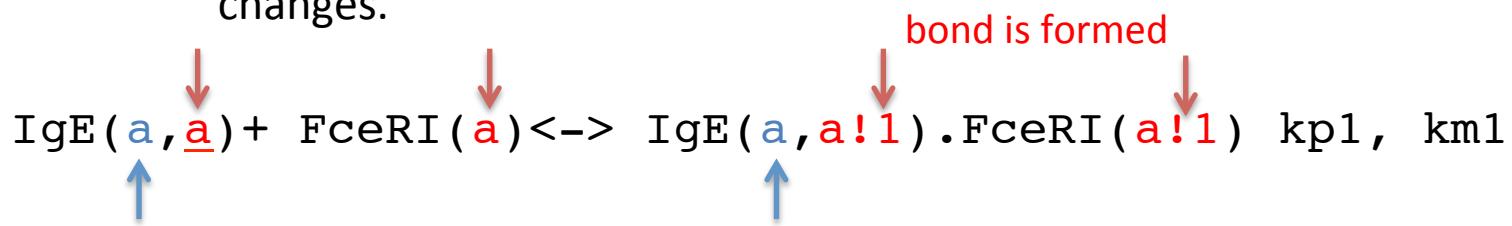
The **center** of a rule is the part that the rule changes.



The **context** is the part that is necessary for the rule to happen but is unchanged.

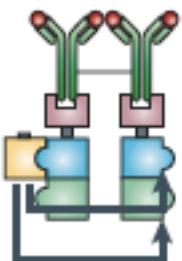
# Center and context

The **center** of a rule is the part that the rule changes.



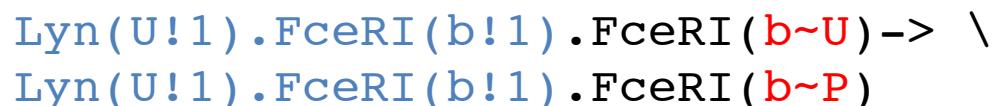
The **context** is the part that is necessary for the rule to happen but is unchanged.

Transphosphorylation



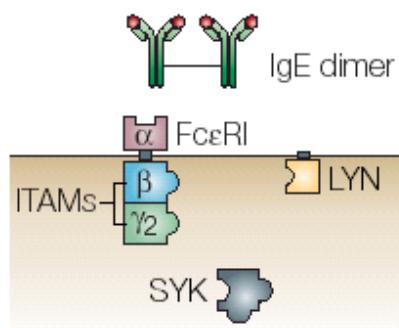
Context can represent complex  
biochemistry.

component state is  
changed

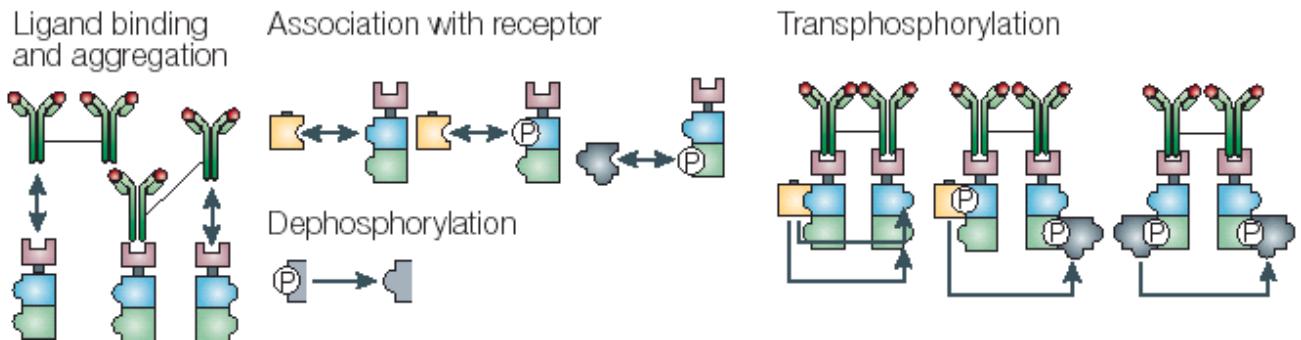


# Composition of a Rule-Based Model

## a Components



## b Interactions



## Molecules

```
begin molecules
Lig(l,1)
Lyn(U,SH2)
Syk(tSH2,l~U~P,a~U~P)
Rec(a,b~U~P,g~U~P)
end molecules
```

## Reaction Rules

```
begin reaction_rules
# Ligand-receptor binding
1 Rec(a) + Lig(l,1) <-> Rec(a!1).Lig(l!1,1) kp1, km1
    Rec(a) + Lig(l,1) <-> Rec(a!1).Lig(l!1,1) kp1, km1

# Receptor-aggregation
2 Rec(a) + Lig(l,1!1) <-> Rec(a!2).Lig(l!2,l!1) kp2, km2

# Constitutive Lyn-receptor binding
3 Rec(b~Y) + Lyn(U,SH2) <-> Rec(b~Y!1).Lyn(U!1,SH2) kpL, kmL
...
```

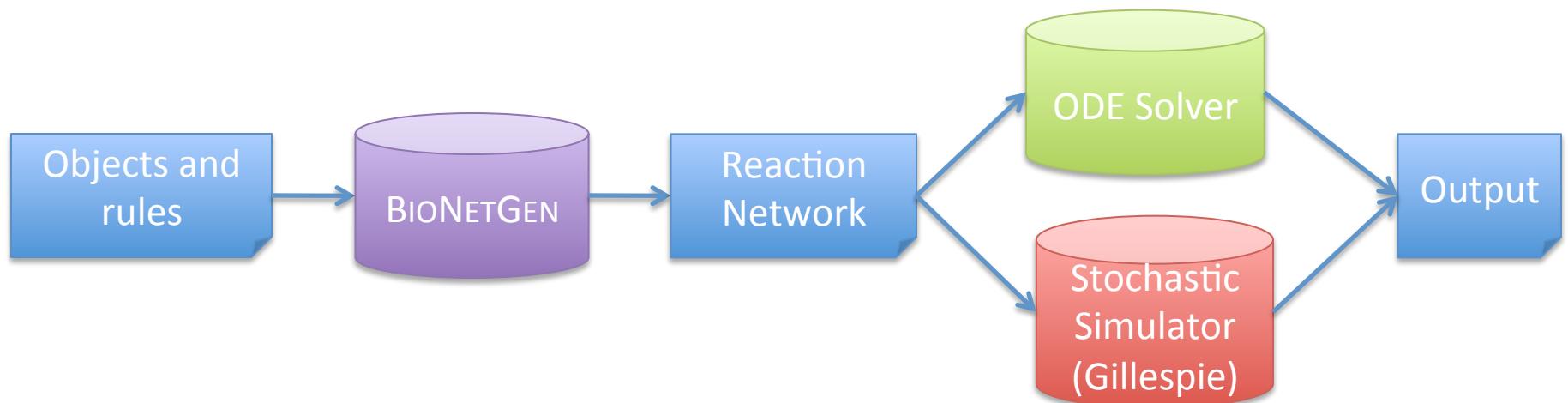
## BioNetGen language

# Applications

- Immunoreceptor Signaling
- Growth factor receptor signaling
- Multivalent binding
- Scaffold effects
- Yeast pheromone signaling
- For a complete list of BioNetGen Applications  
see [http://bionetgen.org/Model\\_Examples](http://bionetgen.org/Model_Examples).

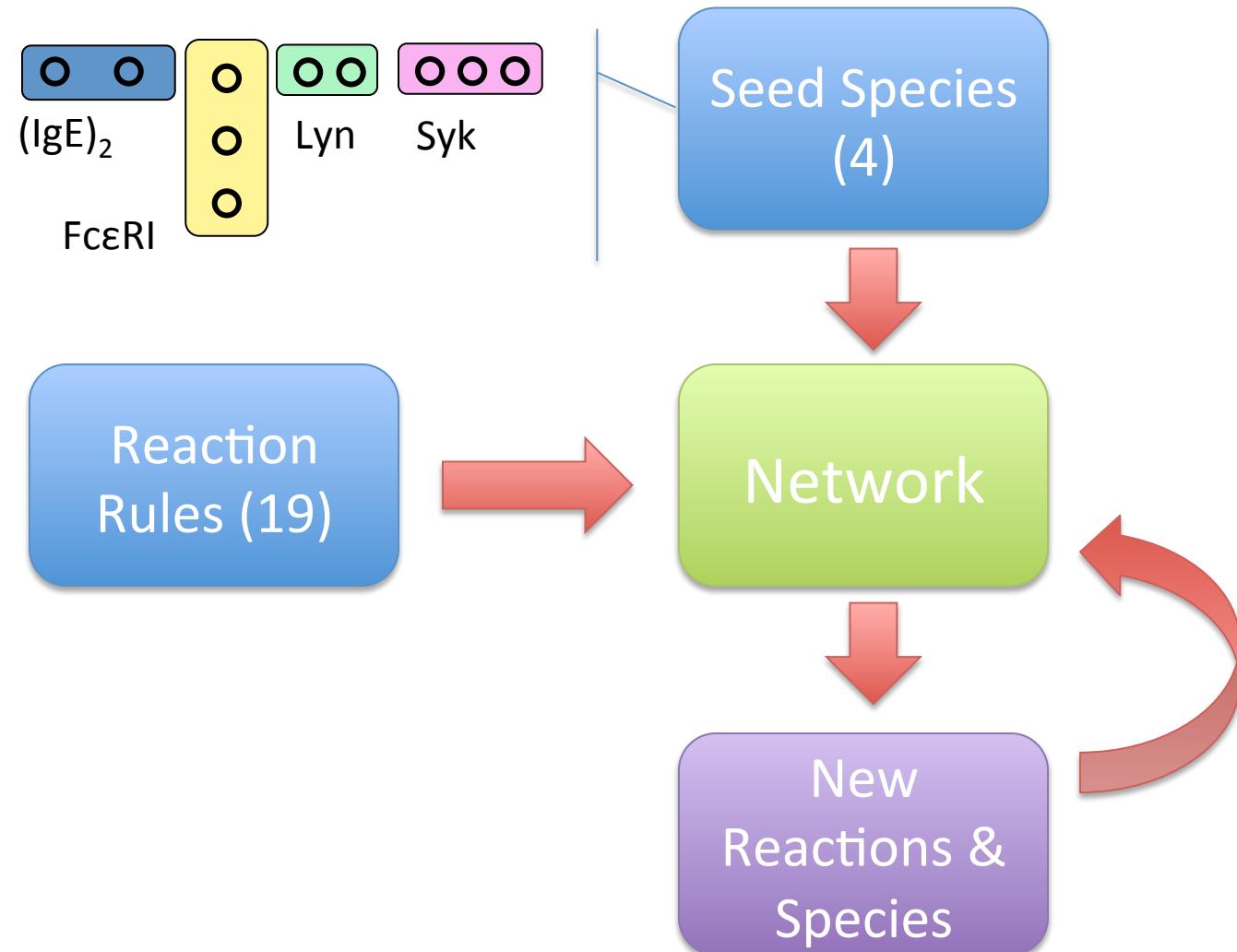
# **SIMULATING A RULE-BASED MODEL**

# Basic RBM workflow with BioNetGen



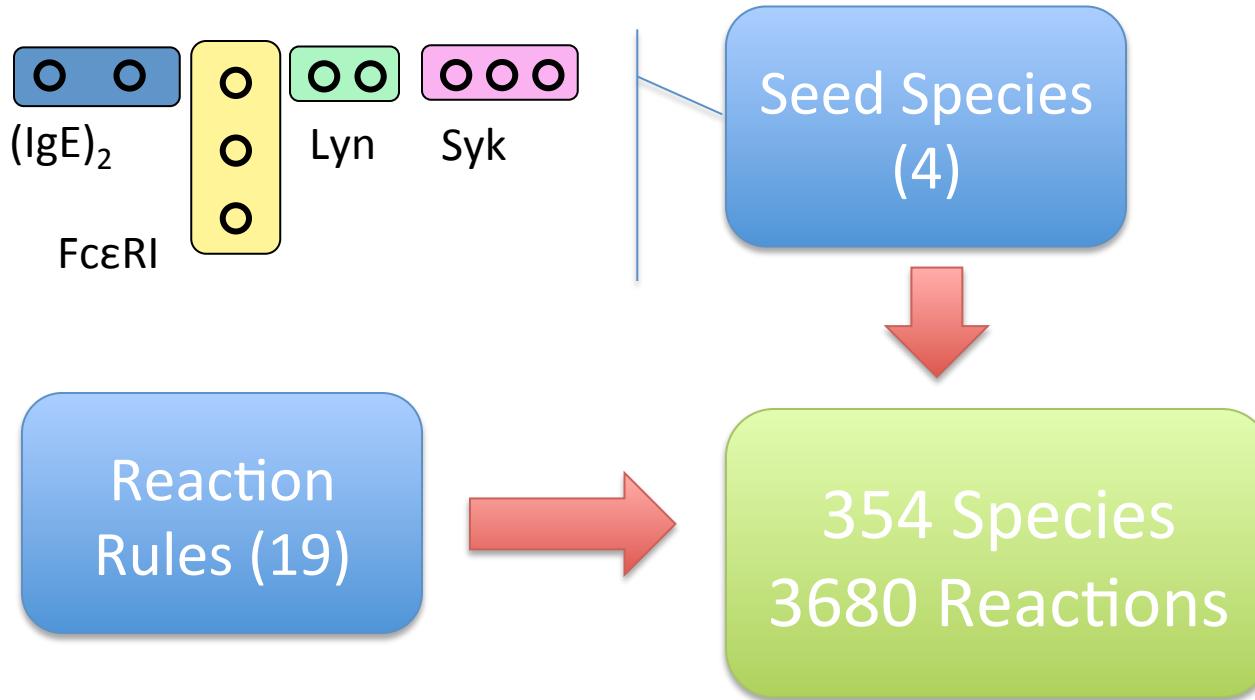
# Automatic Network Generation

## Fc $\epsilon$ RI Model



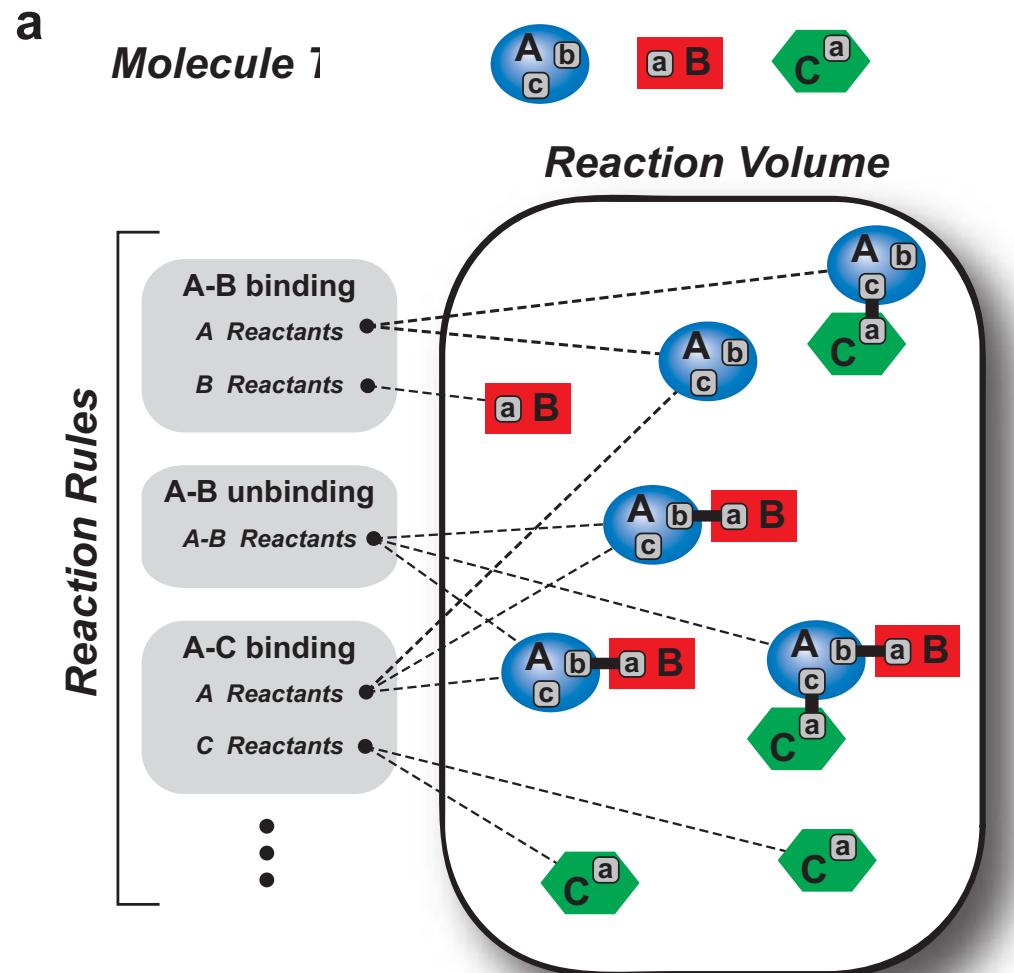
# Automatic Network Generation

## Fc $\epsilon$ RI Model



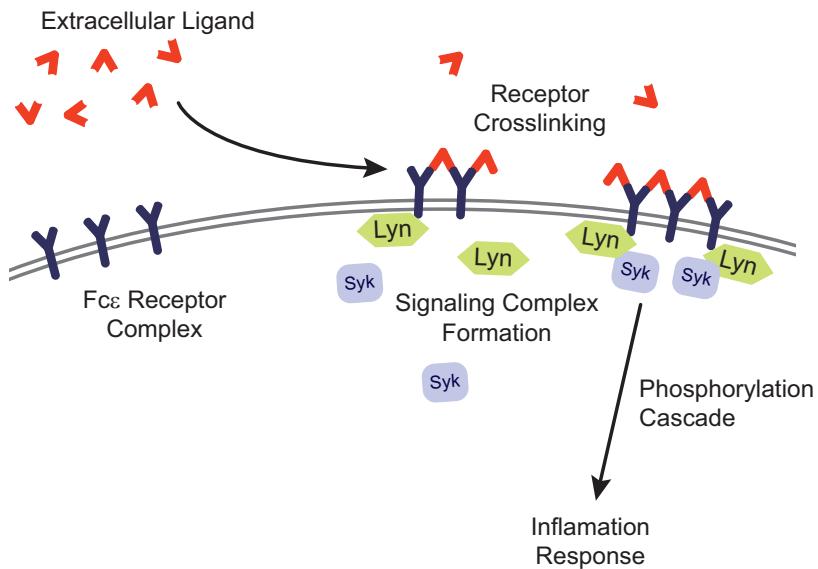
# NFSIM\*

## Network-Free Stochastic Simulator

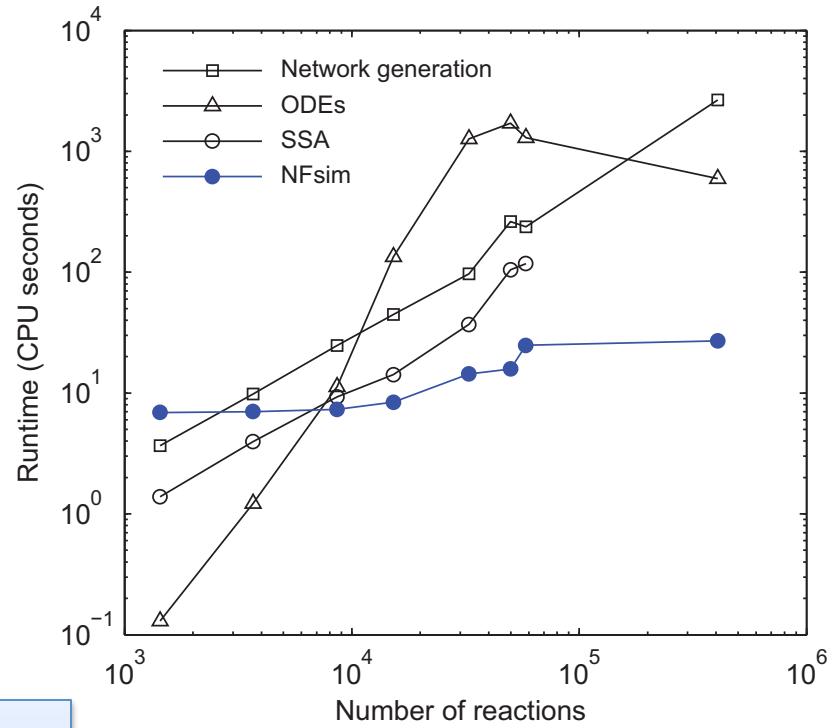


# Fc $\epsilon$ RI signaling models

a



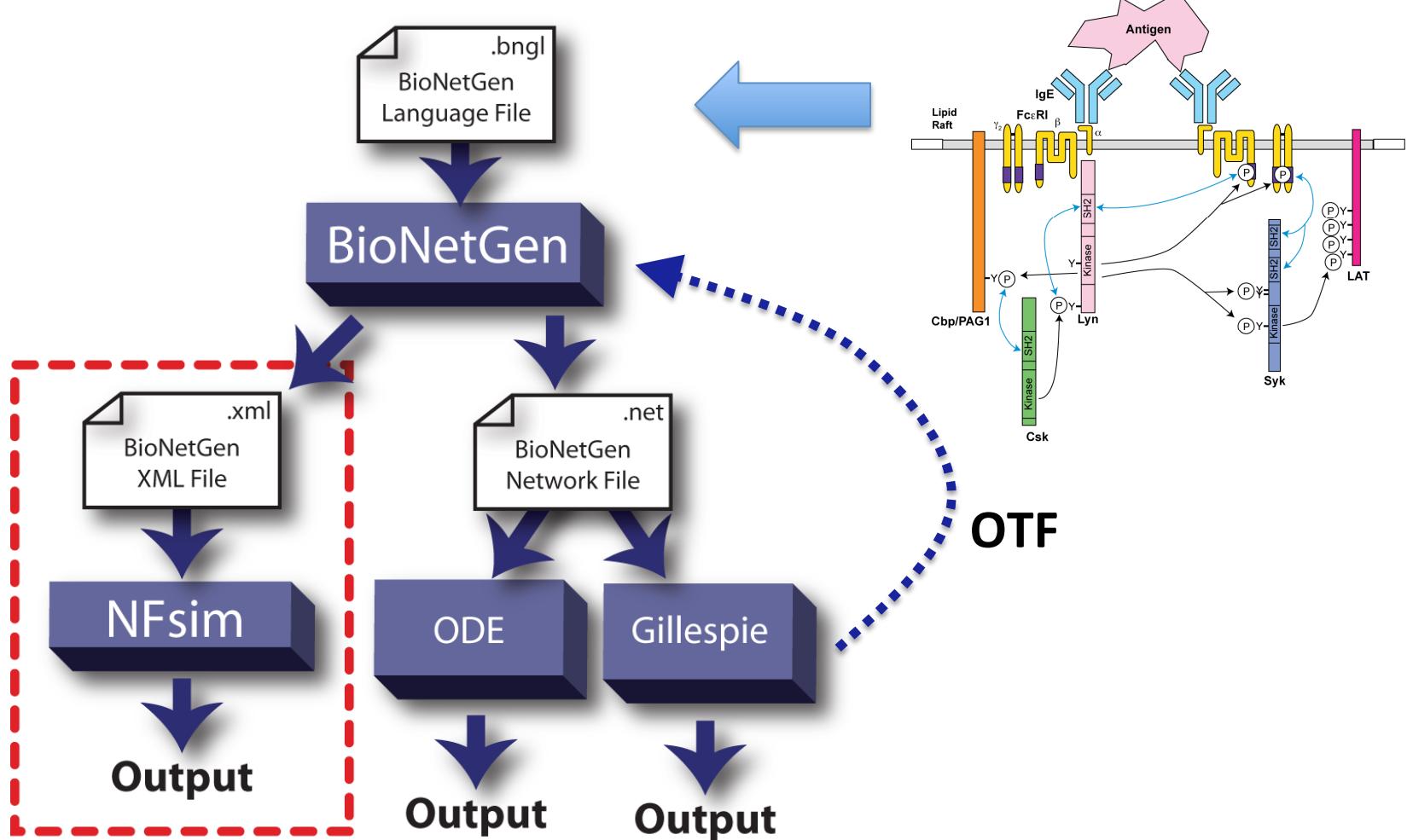
b



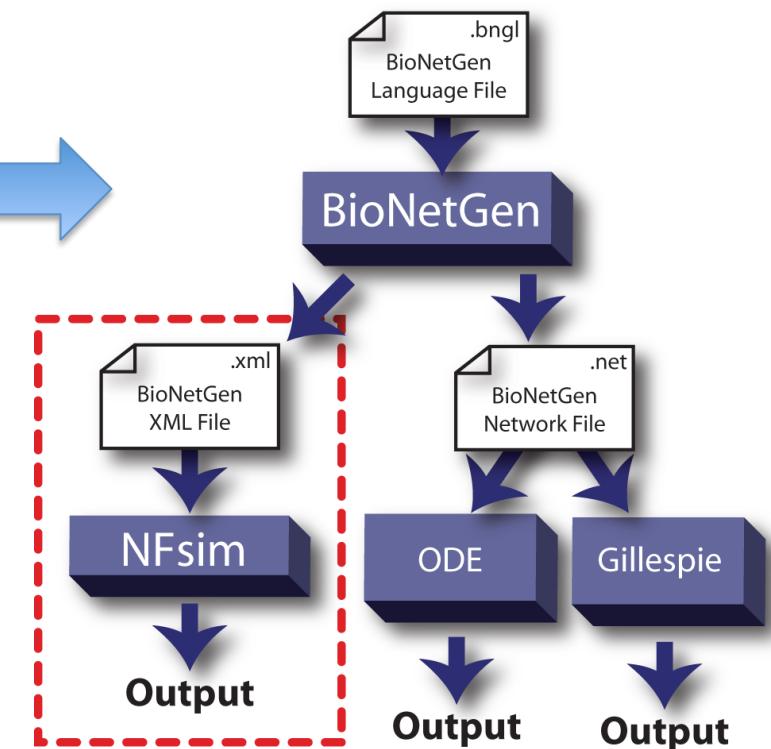
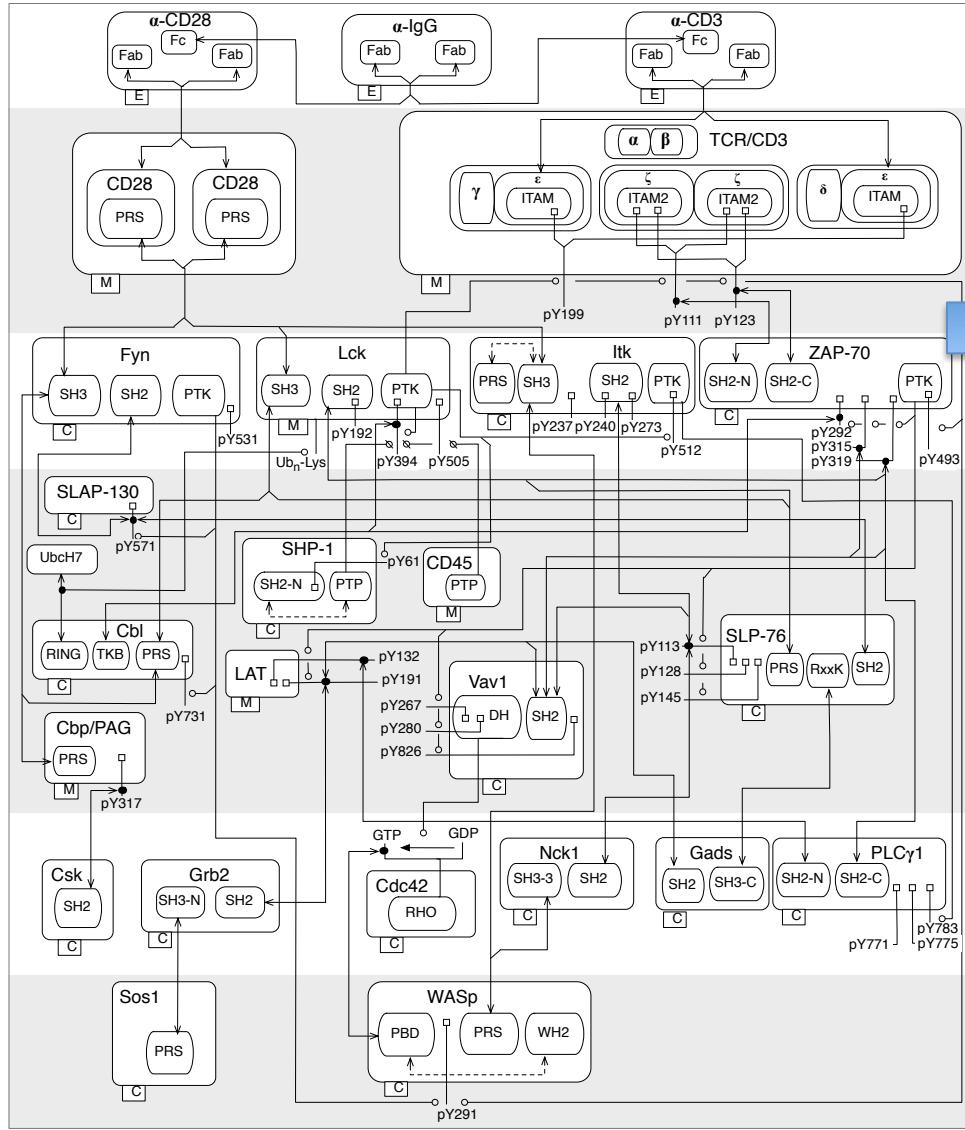
NFSIM can simulate models of greatly increased complexity with manageable increase in cost.

**Increasing complexity**

# Integration with BioNetGEN



# Large Scale TCR Signaling Model



Hu, Chylek, and Hlavacek, unpublished.

# RuleBender

Built in Eclipse RCP

<http://rulebender.org>

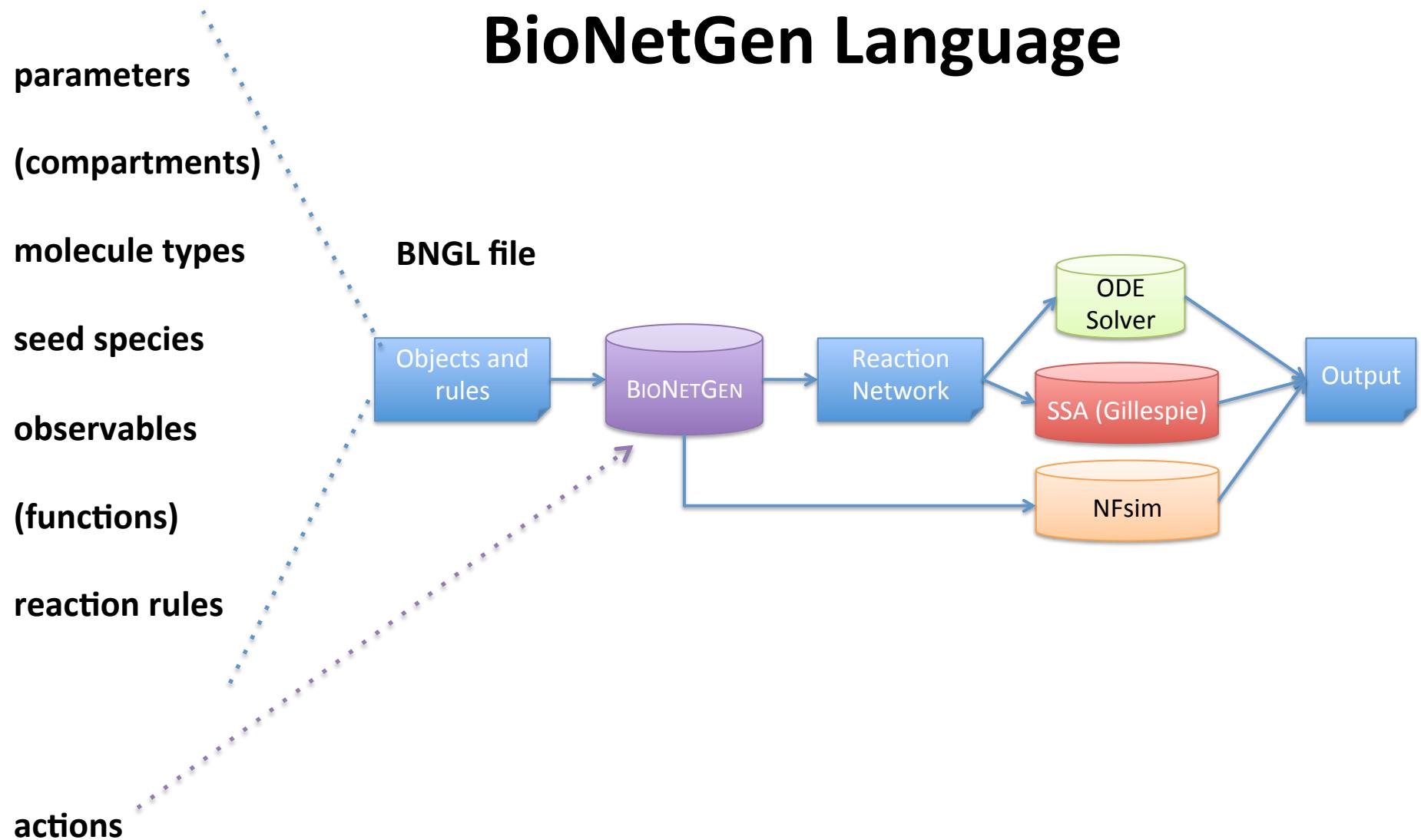
The screenshot shows the RuleBender interface within an Eclipse RCP application window titled "RuleBender".

- Model Editor:** On the left, the "Model" tab is selected, displaying BNGL (Biochemical Notation) code for an EGFR signaling network. The code includes rules for dephosphorylation, transphosphorylation, and activation of Grb2 and Shc proteins.
- Contact Map:** In the center, a "Contact Map" view illustrates the spatial arrangement of proteins and their phosphorylation states. Nodes represent proteins: egfr, Shc, Grb2, PTB, and Sos. States are shown as colored boxes: yellow for unphosphorylated (Y), green for partially phosphorylated (pY), and red for fully phosphorylated (Y~pY). Lines connect nodes to show interactions.
- Properties View:** On the right, the "Properties" view shows the details of the currently selected rule, "Rule11". The rule expression is `egfr(Y1068~pY) + Grb2(SH2,SH3) <-> egfr(Y1068~pY!1).Grb2(SH2!1,SH3)`. The rule label is "Rule11".
- Problems View:** At the bottom left, the "Problems" view indicates "1 error, 0 warnings, 0 others". A detailed table shows the error: "rule parameter\_def failed pre..." at line 17 of the file "egfr\_net.bngl" under resource "/EGFR".

Xu et al. *Bioinformatics* (2011); Smith et al. *BioVis12 (Best Paper)*; BMC Bioinformatics (2012)

# **HANDS-ON TUTORIAL**

# Technical Overview of the BioNetGen Language

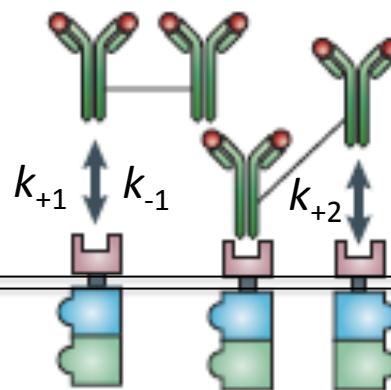


# Dimerization Model

Outer wall (wall)

Ligand binding  
and aggregation

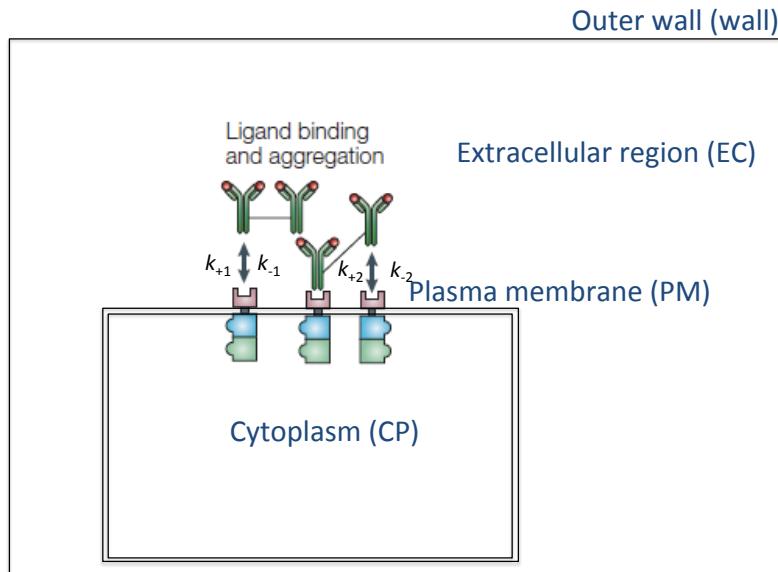
Extracellular region (EC)



Plasma membrane (PM)

Cytoplasm (CP)

# Compartment Specification



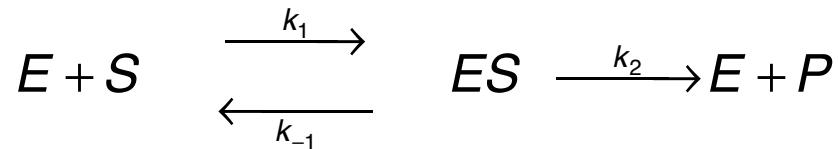
```
begin compartments
wall 2 vol_wall
EC   3 vol_EC    wall
PM   2 vol_PM    EC
CP   3 vol_CP    PM
end compartments
```

Volume of surface compartment = Area\*thickness  
thickness = 10 nm = 0.01  $\mu\text{m}$

# **BACKUP EXAMPLE**

# Example 1: MM Mechanism

parameters



molecule types

seed species

A BioNetGen model consists of a set of blocks, each beginning and ending with `begin <blockname>` / `end <blockname>` respectively.

observables

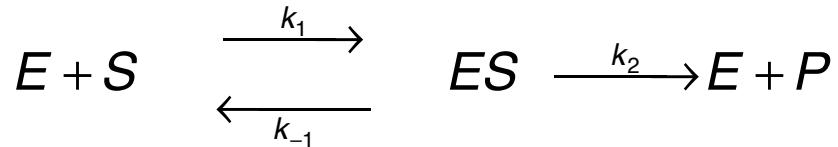
functions

reaction rules

actions

# Example 1: MM Mechanism

**parameters**



molecule types

seed species

parameters – model constants are defined here. *The user is responsible for using a consistent set of units, which should be indicated in the associated comments.*

observables

functions

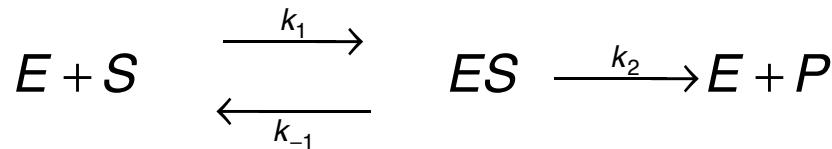
reaction rules

actions

# Example 1: MM Mechanism

**parameters**

molecule types



seed species

observables

functions

reaction rules

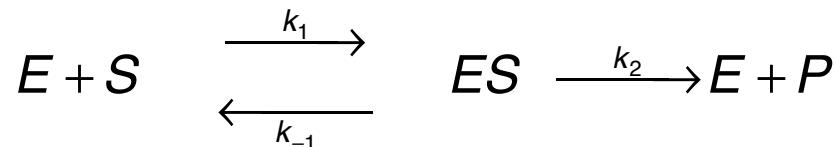
actions

```
begin parameters
    # Avogadro's number- scaled for umol
    NA 6.02e23/1e6
    # Cell volume
    V 1e-12 # liters - typical for eukaryote
    # Rate constants
    kp1  1.0/(NA*V) # 1/uM 1/s-> 1/molec 1/s
    km1  1.0e-1 # 1/s
    k2   1.0e-2 # 1/s

    # Initial concentrations
    E0 0.01*NA*V # uM -> molec / cell
    S0 1.0*NA*V # uM -> molec / cell
end parameters
```

# Example 1: MM Mechanism

parameters



molecule types

molecule types— molecules, their components, and their allowed component states are declared here.

seed species

observables

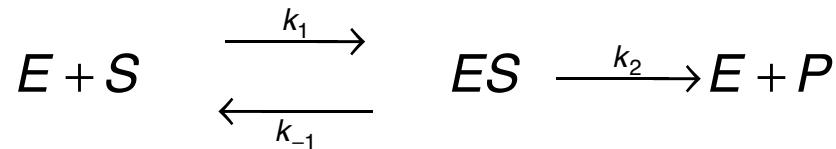
functions

reaction rules

actions

# Example 1: MM Mechanism

parameters



molecule types

seed species

```
begin molecule types
  E(s)
  S(Y~O~P)
end molecule types
```

observables

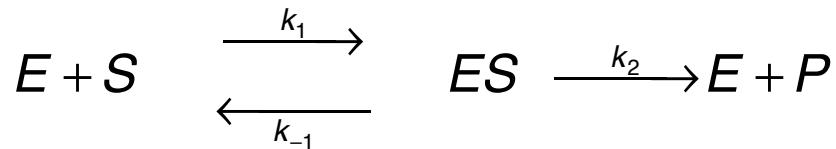
functions

reaction rules

actions

# Example 1: MM Mechanism

parameters



molecule types

**seed species**

seed species— species initially present in the system at time t=0 followed by their initial concentration. Standard is all molecule types in their “ground state” with basal expression levels. May include complexes. All components of molecules that have states must be in a specified state. All complexes must be connected.

observables

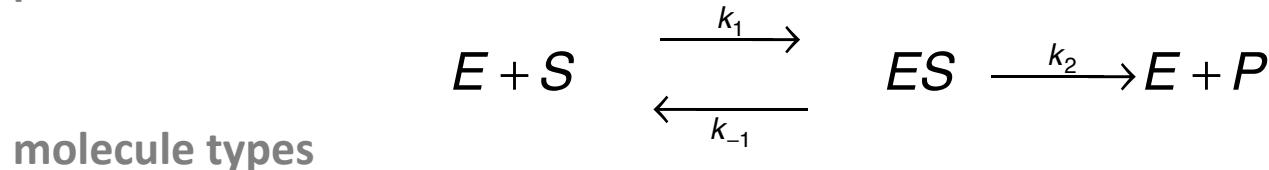
functions

reaction rules

actions

# Example 1: MM Mechanism

parameters



seed species

observables

functions

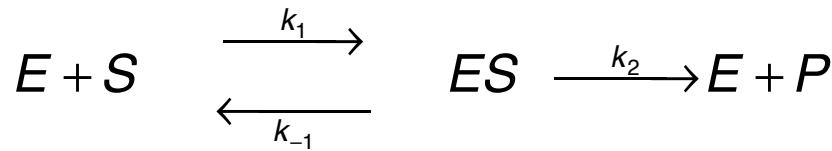
```
begin seed species
    E(s)    E0
    S(Y~0)  S0
end seed species
```

reaction rules

actions

# Example 1: MM Mechanism

parameters



molecule types

seed species

**observables**

observables— Defined sums of concentrations of species with specified properties. Syntax is <type> <name> <pattern>. Types considered here are Molecules and Species, which indicate weighted and unweighted sums respectively. These are used to define model outputs and are used as to make the default plot in RuleBender.

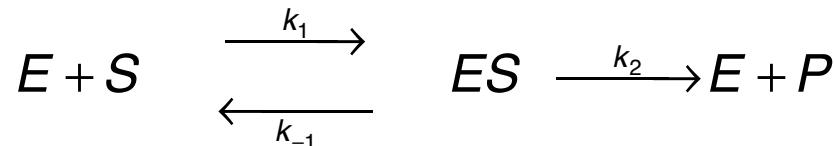
functions

reaction rules

actions

# Example 1: MM Mechanism

parameters



molecule types

seed species

```
begin observables
Molecules SU S(Y~0)
Molecules SP S(Y~P)
Molecules ES E(s!1).S(Y!1)
end observables
```

observables

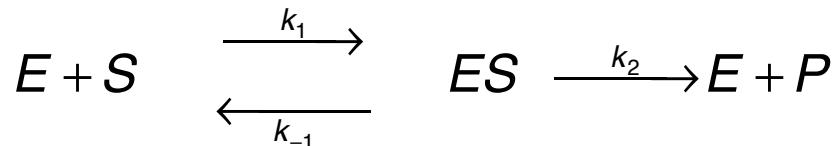
functions

reaction rules

actions

# Example 1: MM Mechanism

parameters



molecule types

seed species

```
begin observables
Molecules SU S(Y~0)
Molecules SP S(Y~P)
Molecules ES E(s!1).S(Y!1)
end observables
```

observables

functions

observable	SU	S(Y~0)	S(Y~0)
matches			
species	S(Y~0)	E(s!1).S(Y~0!1)	

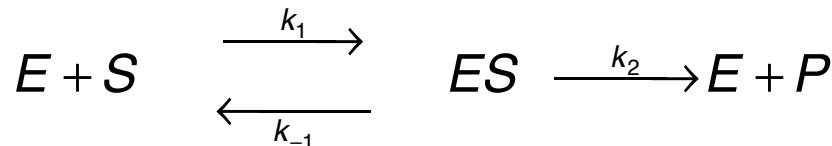
reaction rules

$SU = \text{sum of concentration of matches} = [S(Y~0)]$

actions

# Example 1: MM Mechanism

parameters



molecule types

seed species

```
begin observables
Molecules SU S(Y~0)
Molecules SP S(Y~P)
Molecules ES E(s!1).S(Y!1)
end observables
```

observables

functions

observable ES       $E(s!1).S(Y!1)$   
matches  
species             $E(s!1).S(Y\sim 0!1)$

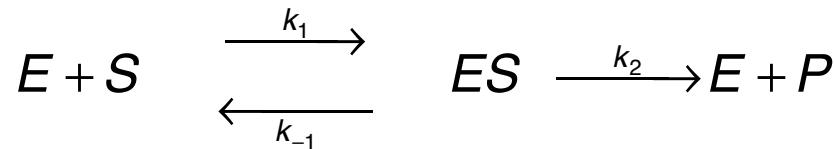
reaction rules

actions

$$ES = \text{sum of concentration of matches} = [E(s!1).S(Y\sim 0!1)]$$

# Example 1: MM Mechanism

parameters



molecule types

seed species

observables

functions

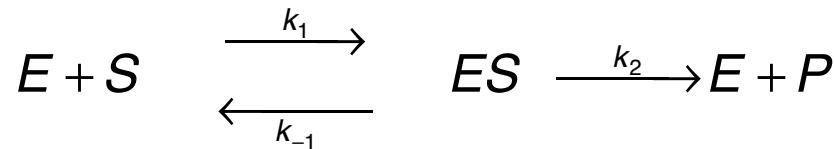
**reaction rules**

actions

reaction rules— Rules that generate reactions based on selecting reactants with specified properties and transforming them in a specified way with the specified rate law. Syntax is <name>: <reactants> <arrow> <products> <rate law>. Name is optional but useful.

# Example 1: MM Mechanism

parameters



molecule types

seed species

```
begin reaction rules
```

observables

```
ESbind: \
E(s) + S(Y~0) <-> E(s!1).S(Y~0!1) kp1, km1
```

functions

```
ESconvert: \
E(s!1).S(Y~0!1) -> E(s) + S(Y~P) k2
```

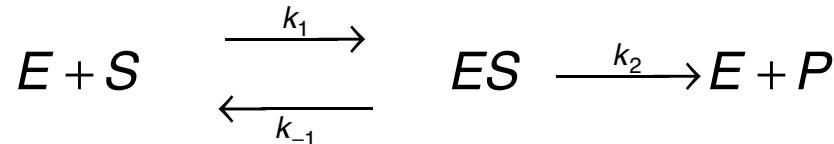
reaction rules

```
end reaction rules
```

actions

# Example 1: MM Mechanism

parameters



molecule types

actions– Need not be enclosed in block. Come after model definition and specify simulation protocol for a model.

seed species

```
generate_network({});  
simulate_ode({t_end=>1000, n_steps=>100});
```

observables

functions

reaction rules

actions