



Mechanism of chloride channeling by Excitatory Amino Acid Transporters

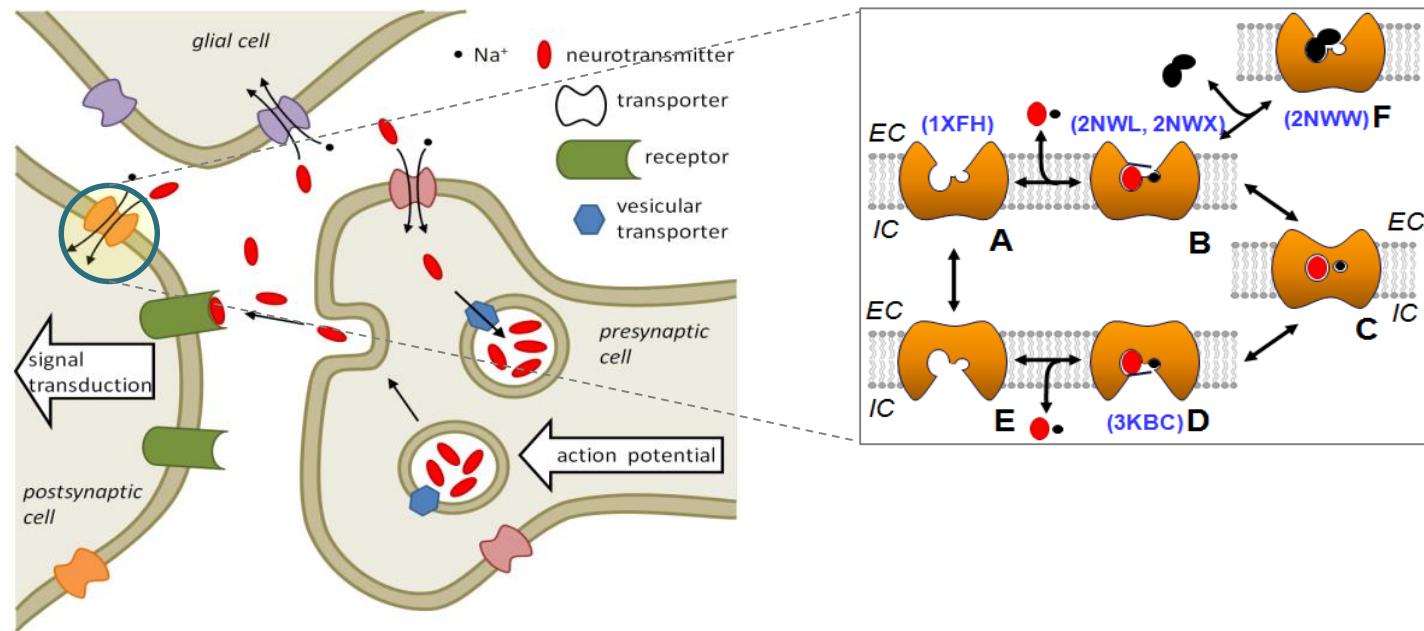
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School of Medicine, University of Pittsburgh*



Susan Amara

GLUTAMATERGIC SIGNALING



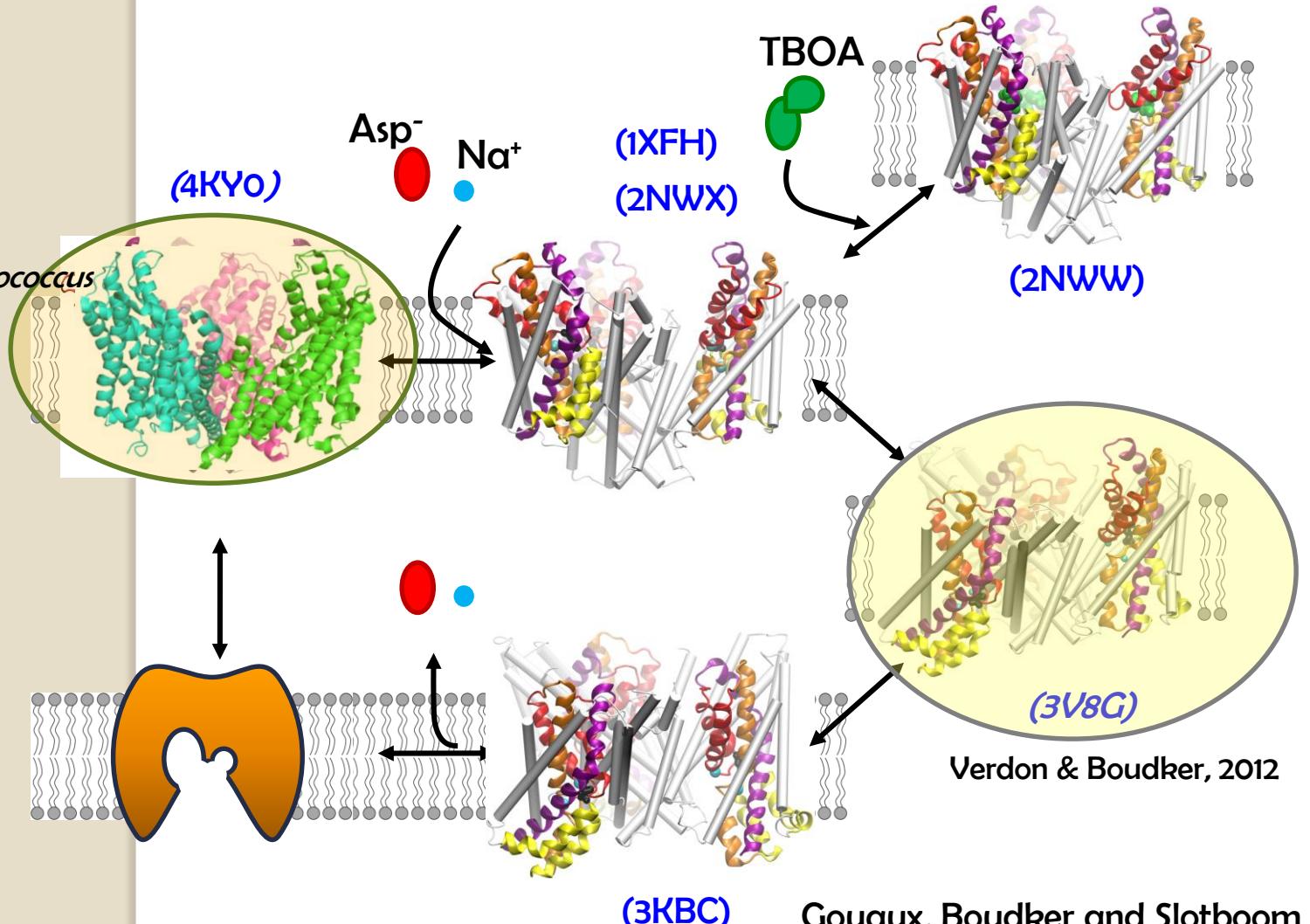
References

Gu et al (2009) *Proc Natl Acad Sci USA* 106: 2589-94; Shrivastava et al (2008) *J Biol Chem* 283: 28680-90

EAAT'S ALTERNATING ACCESS MECHANISM

Jensen et al., 2013

GltTk from *Thermococcus kodakarensis*

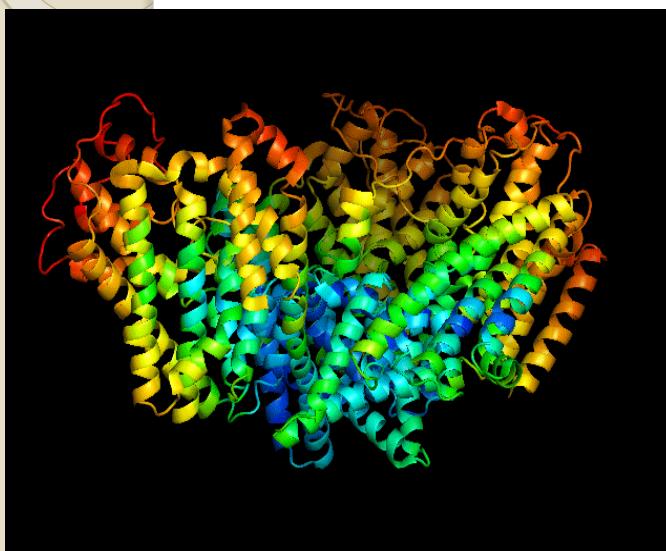


References

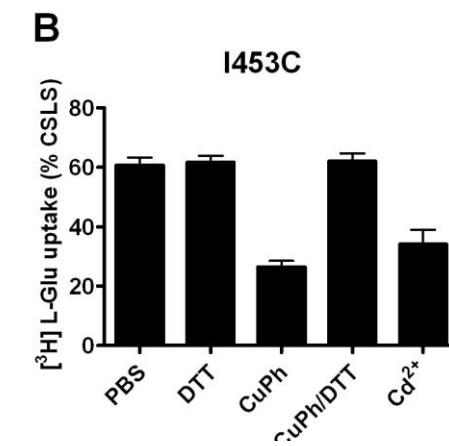
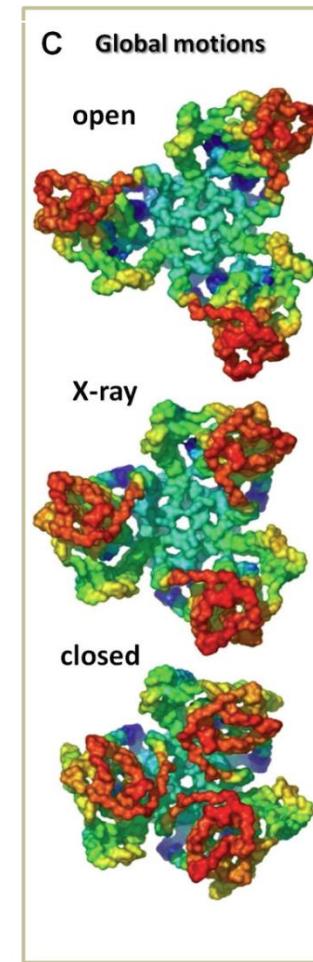
Lezon & Bahar (2012) *Biophys J* 102: 1331-40; Jiang et al (2011) *Proc Natl Acad Sci USA* 108: 15141-6

DeChancie et al (2011) *Mol. BioSyst* 7:832-42;

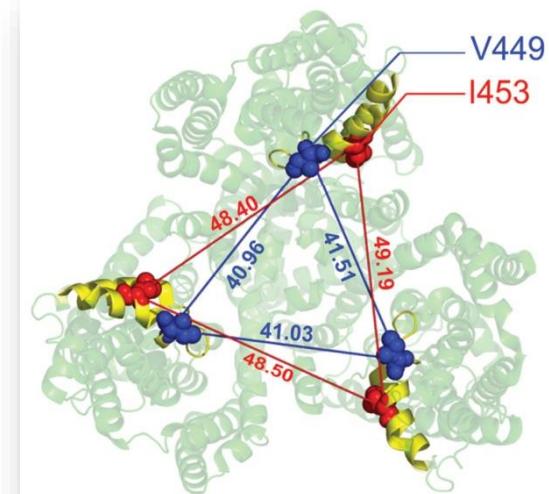
Cooperative Intersubunit Motions are Essential to Function



Jiang et al., PNAS 2011

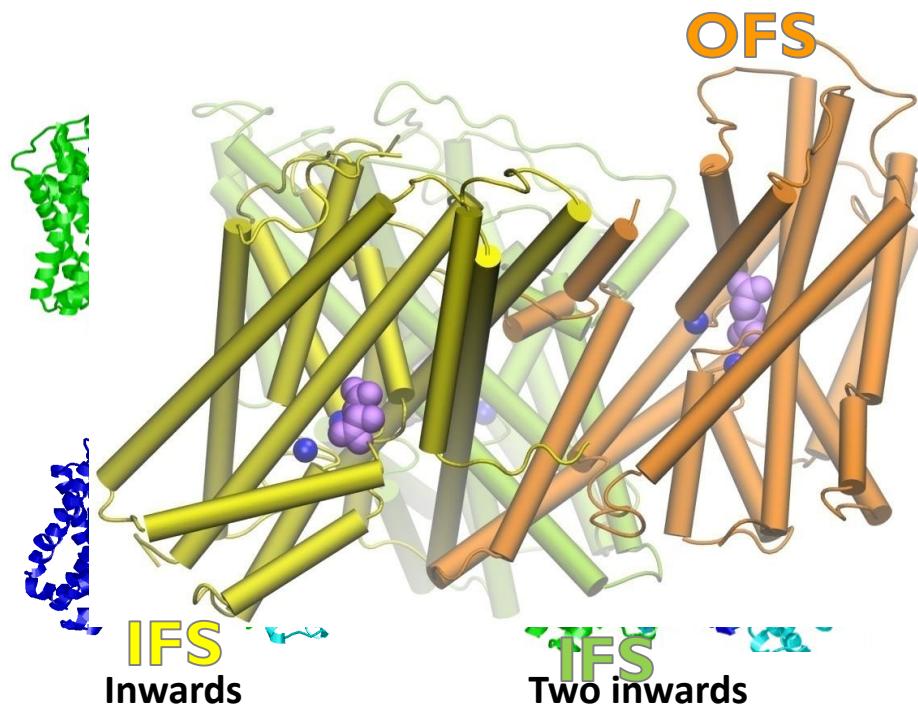
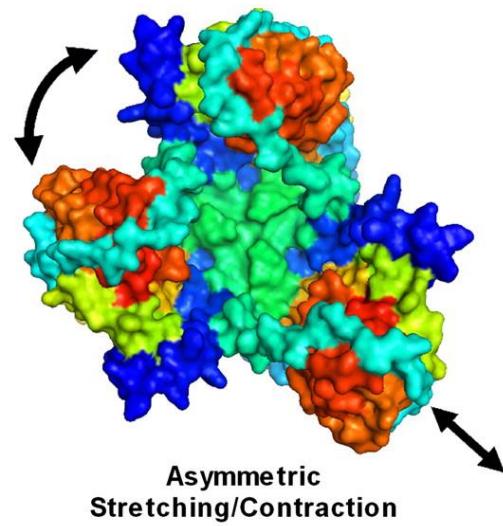


Residues mapped onto EAAT1

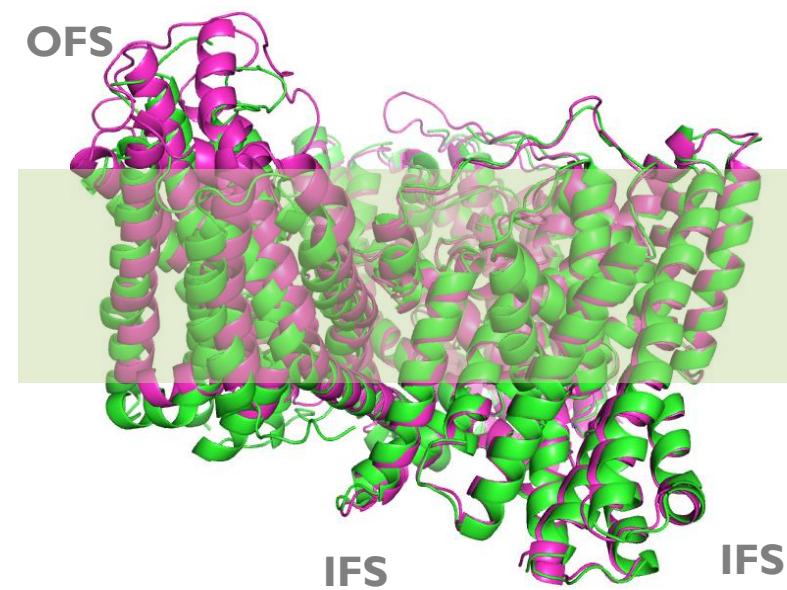
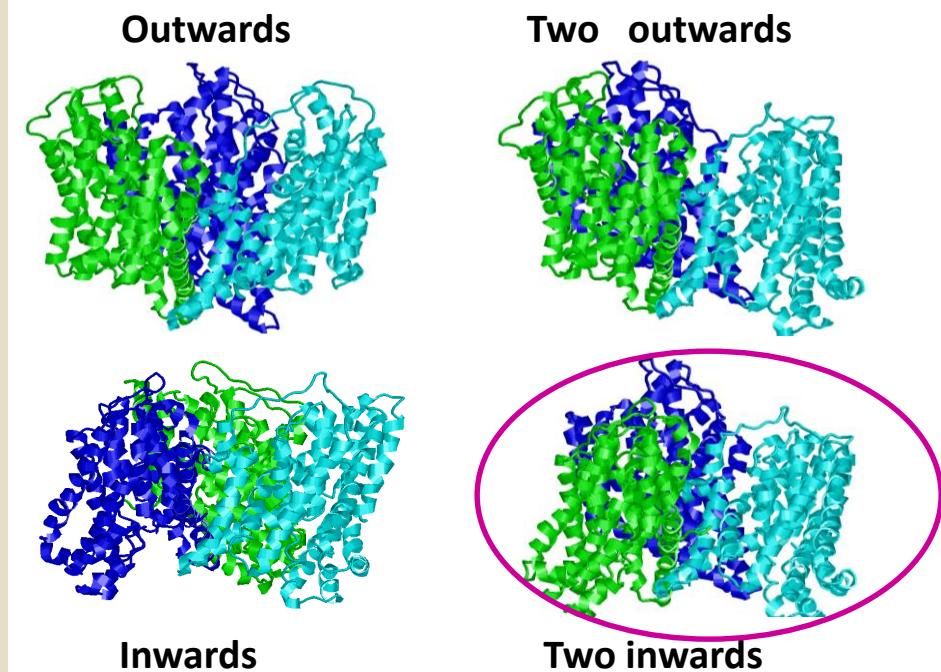


Glutamate uptake blocked by disulfide cross-linking between 'distant' pairs

Transition of one subunit at a time: a mechanism revealed by our experiments & computations

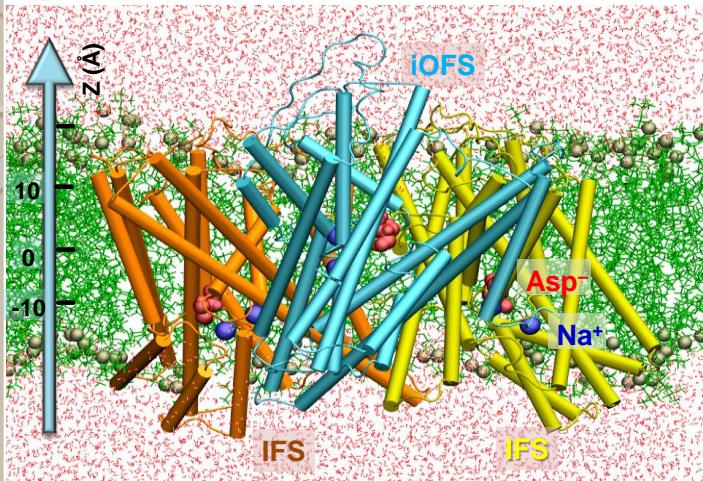


Transition of one subunit at a time: a mechanism revealed by our experiments & computations

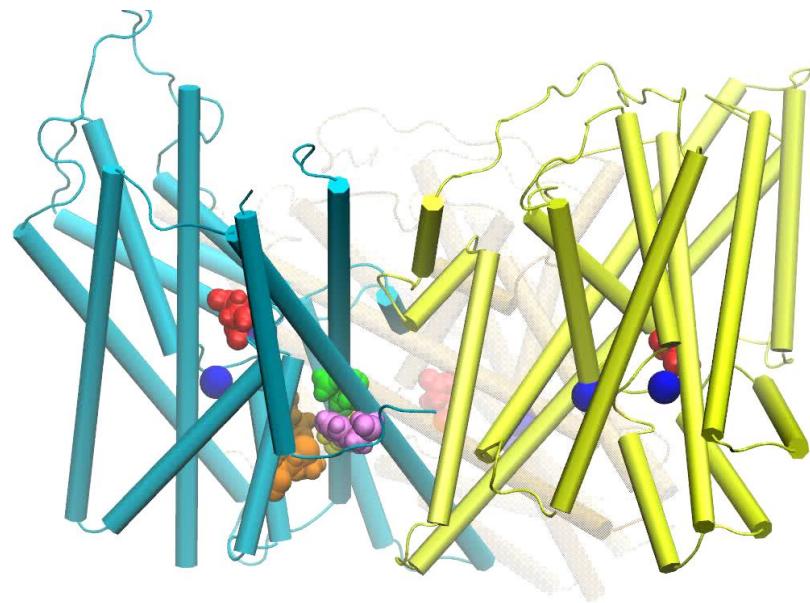
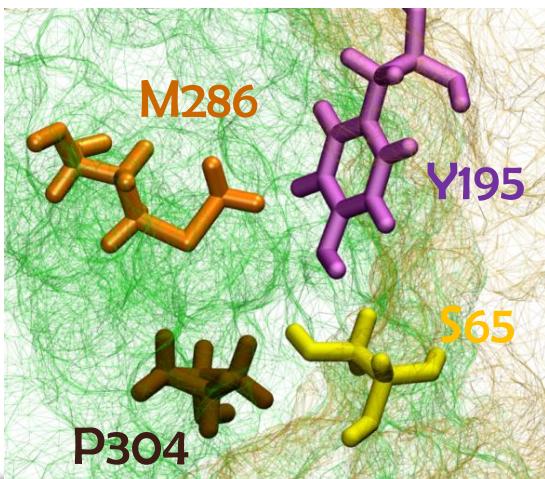


$$\text{RMSD} = 1.382 \text{ \AA}$$

Intermediates mediate permeation of polar solutes

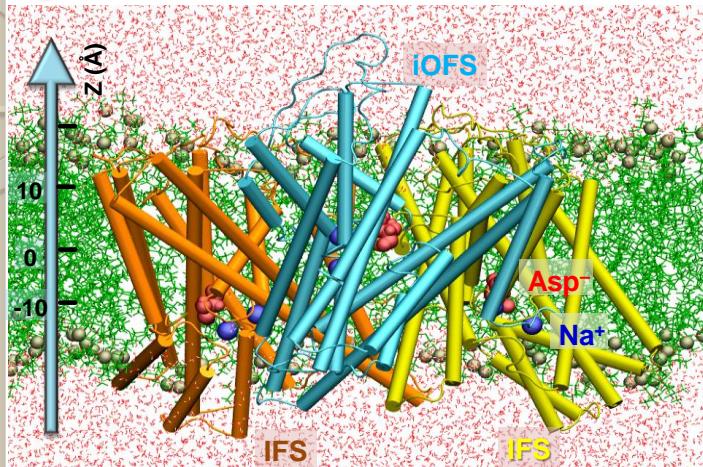


MD setup for Glt_{Ph} (PDB:3V8G)

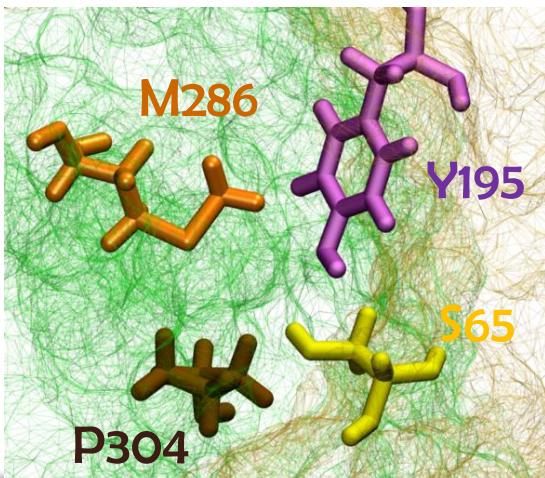
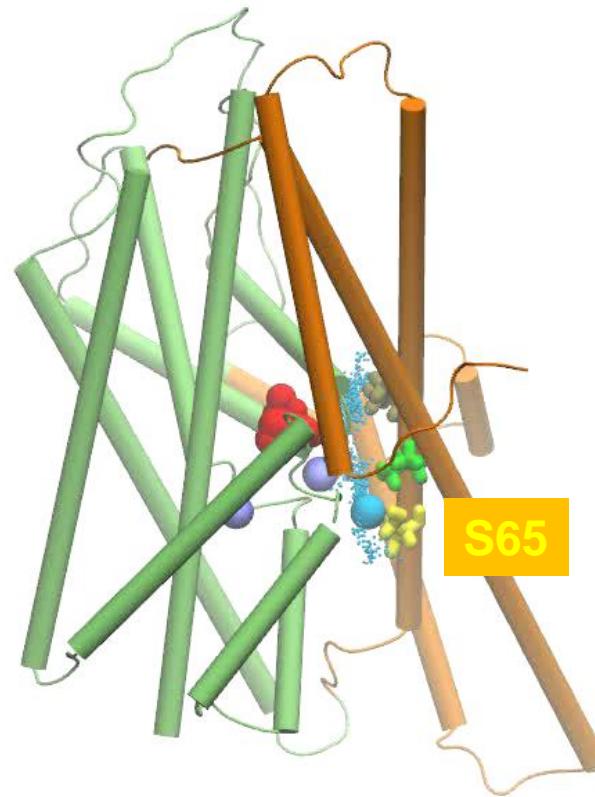


The putative anion pathway suggested by Verdon and Boudker shows a constriction zone near S65, Y195, M286 and P304

Intermediates mediate permeation of polar solutes

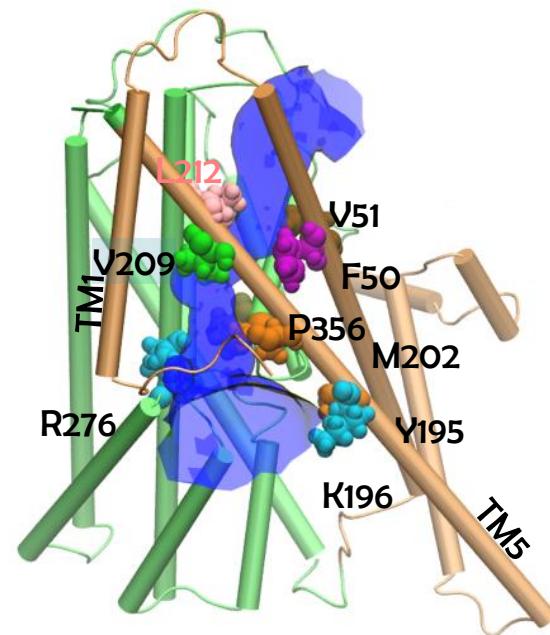
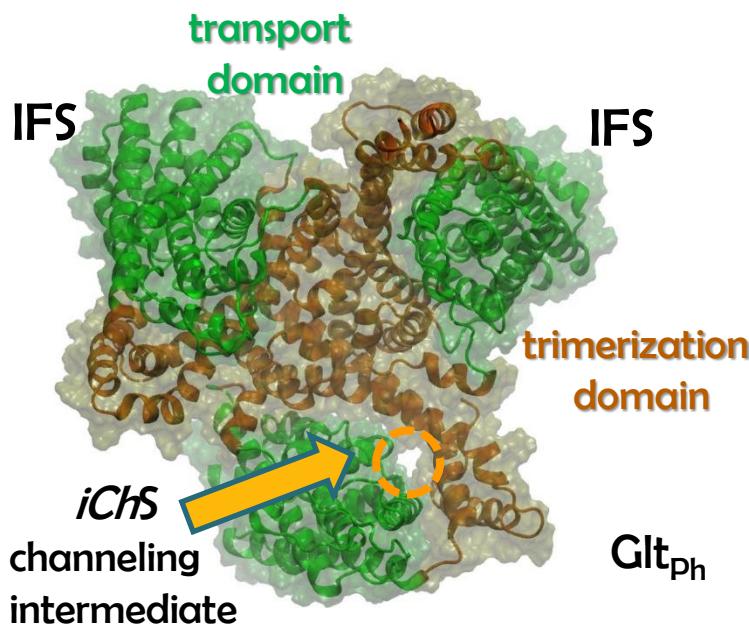


MD setup for Glt_{Ph} (PDB:3V8G)



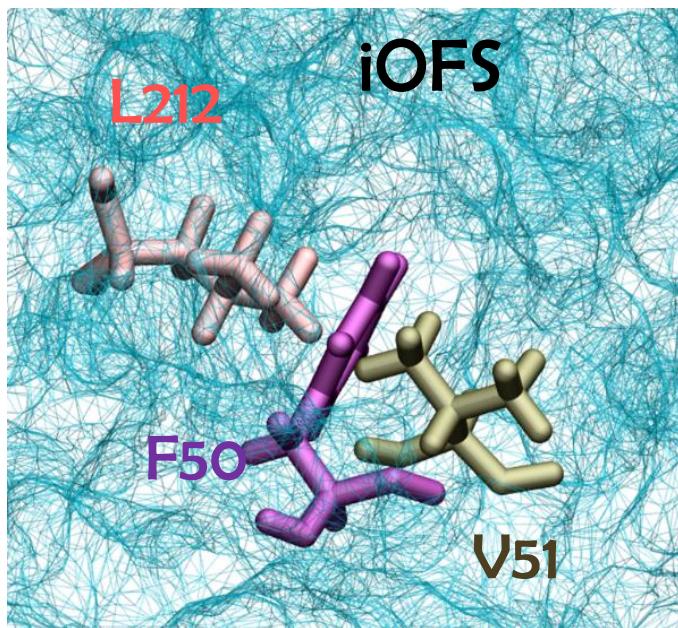
Energy barrier ~20 kcal/mol

A intermediate water-channeling state iChS forms during the transition to inward-facing conformations

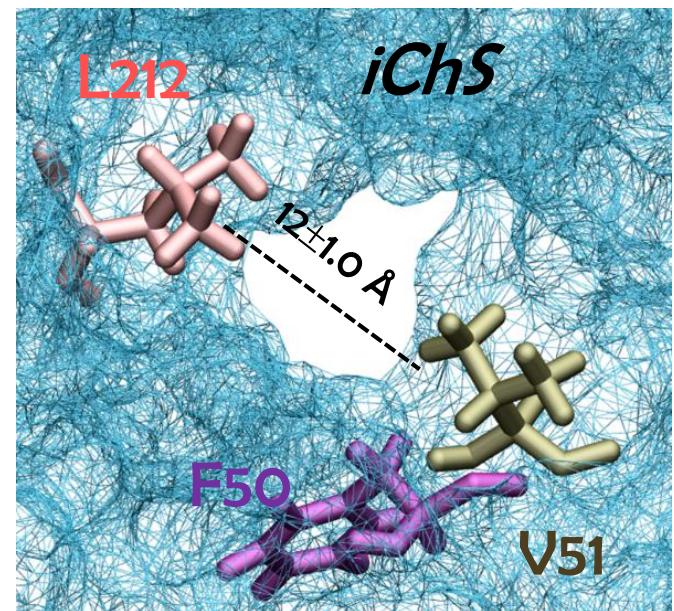


A intermediate water-channeling state iChS forms during the transition to inward-facing conformations

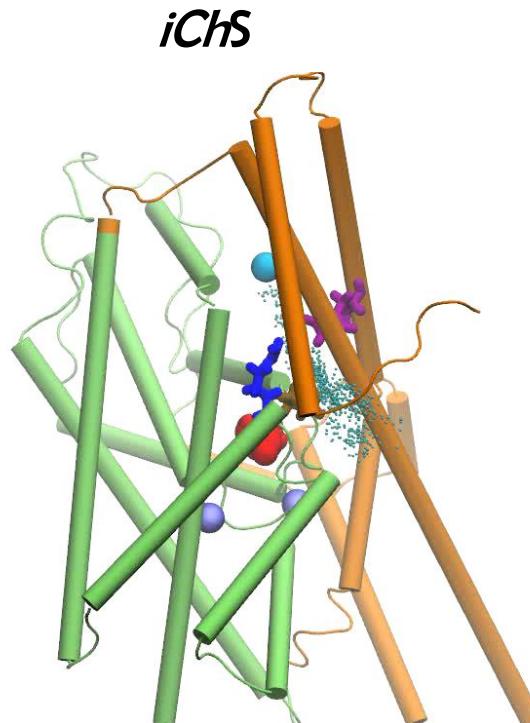
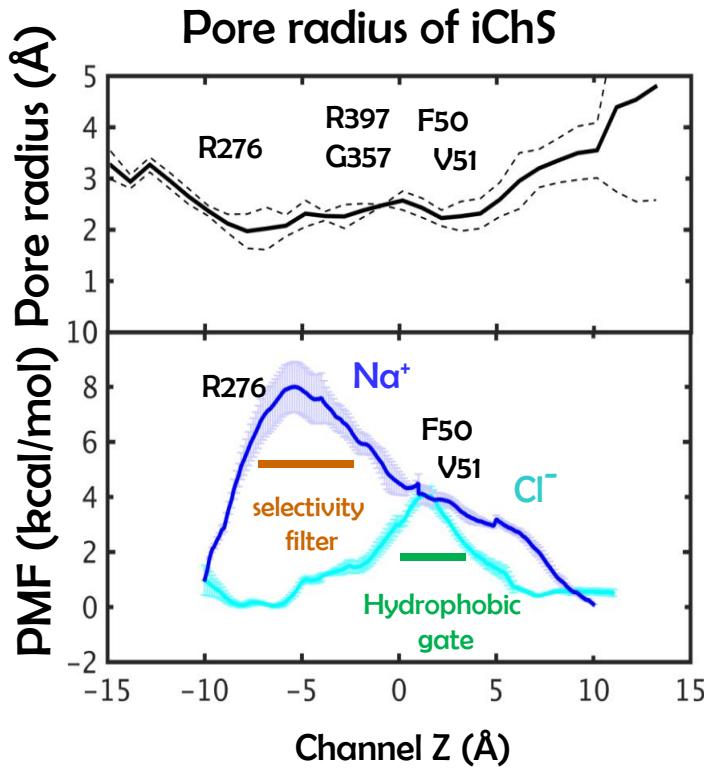
Closed state



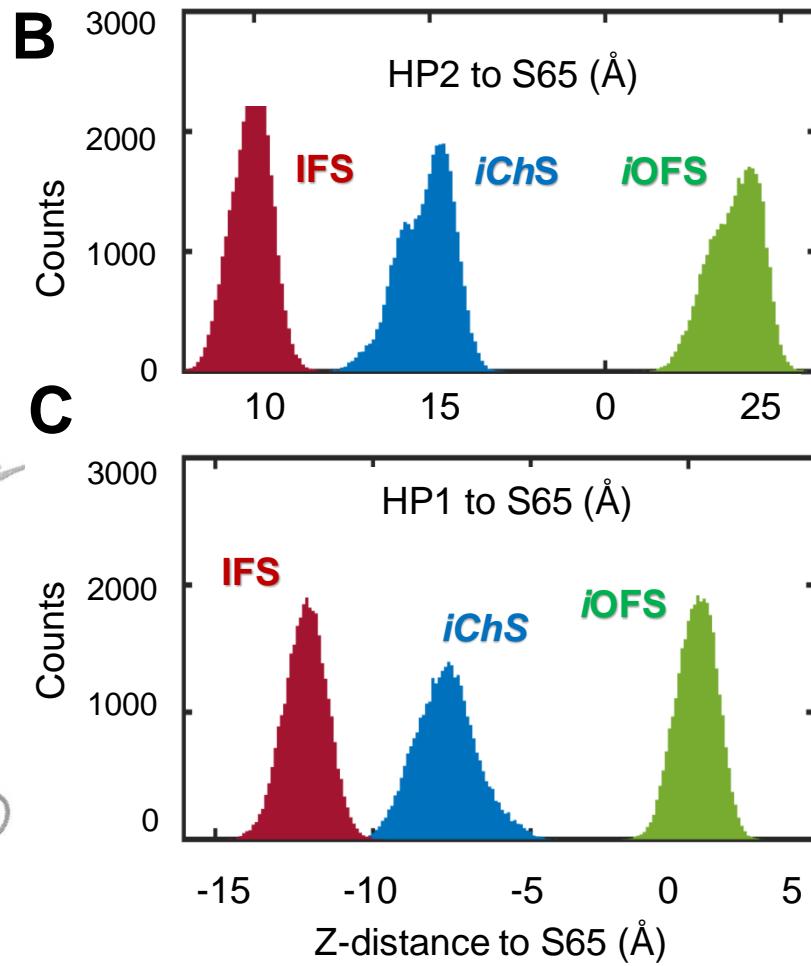
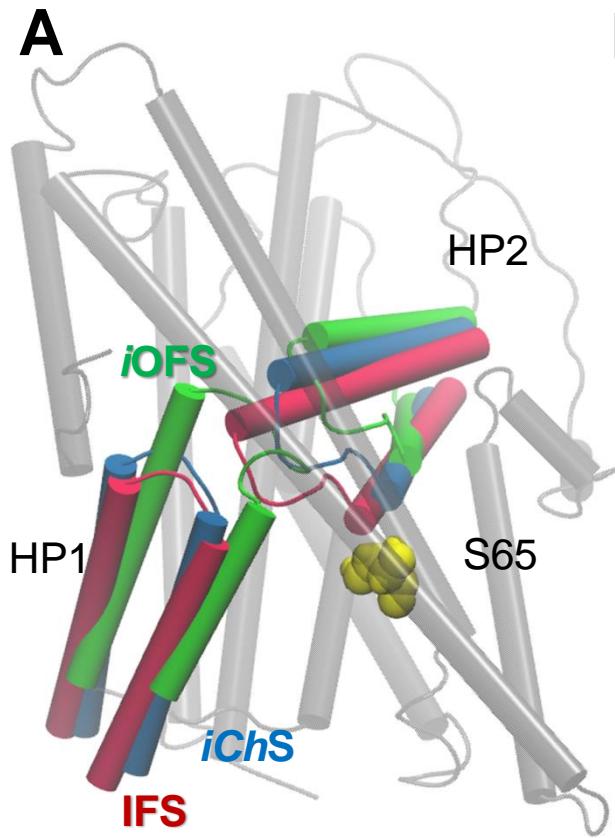
Open state



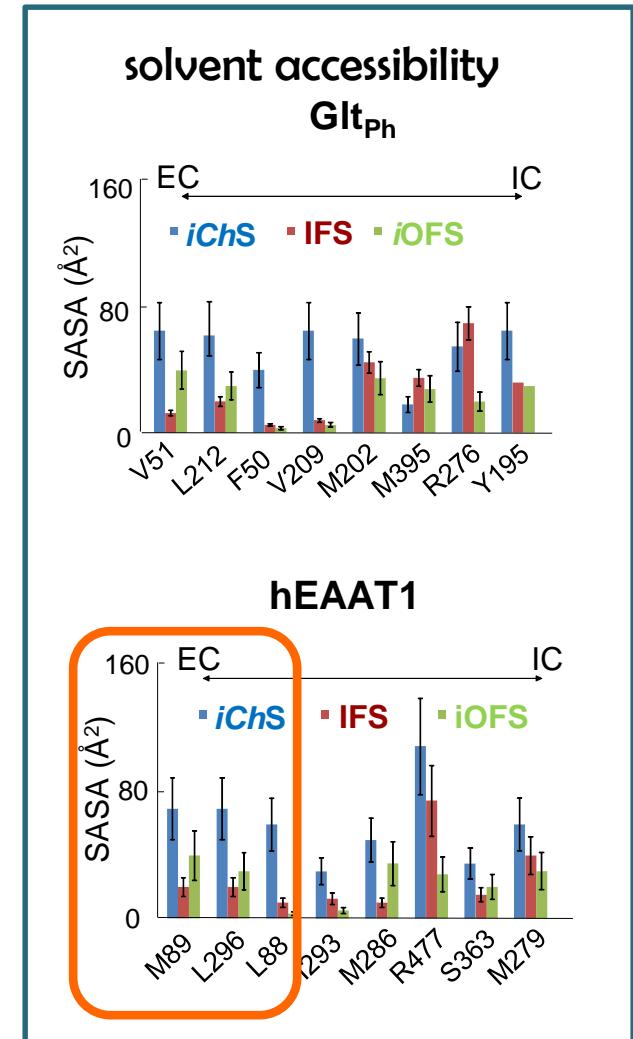
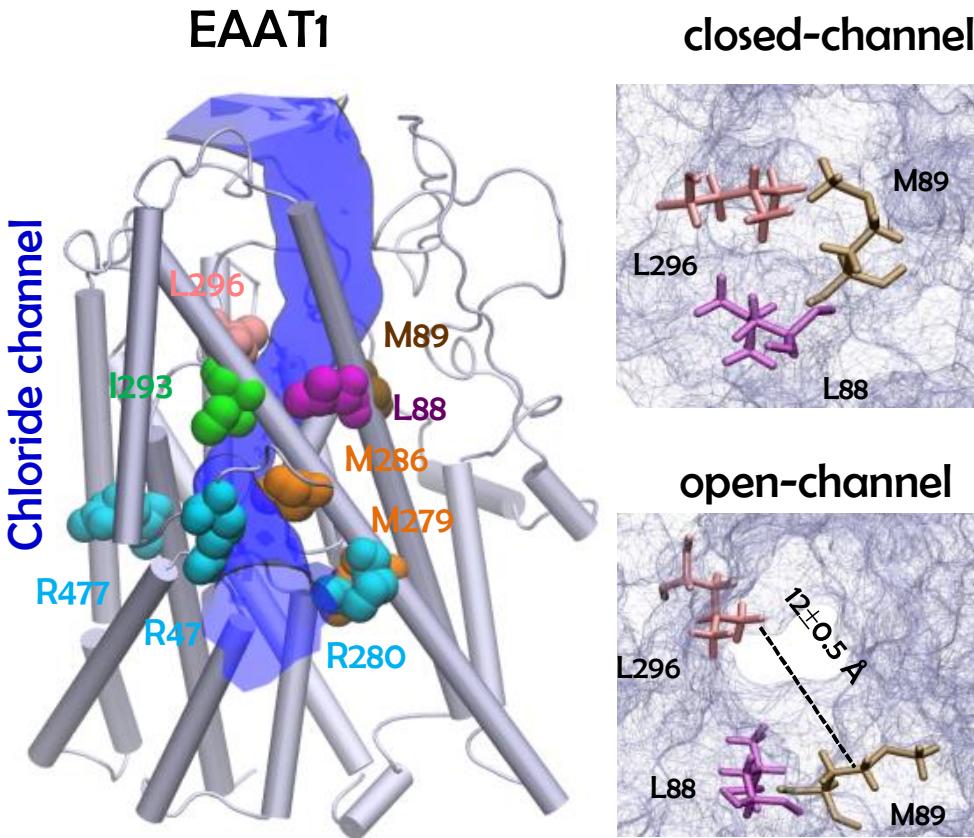
iChS favors anion permeation



iChS distinct from OF and IF states



Same channel confirmed in EAAT1



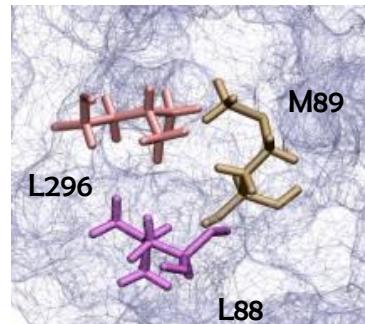
Hypothesis:

Three residues are found to control the channel opening or closure, and they are solvent-exposed prior to transitioning to closed state.

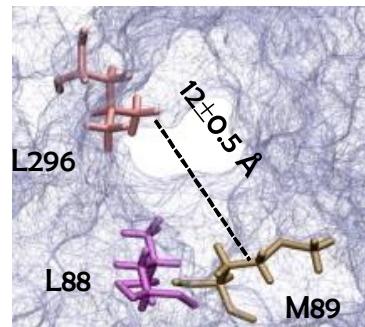
Suppose we bind at those positions a bulky molecule at those positions.

Do we observe an effect on channel permeability?

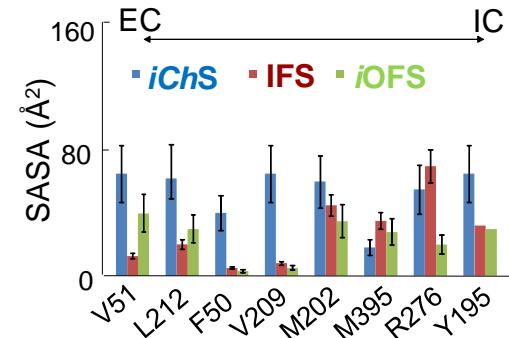
closed-channel



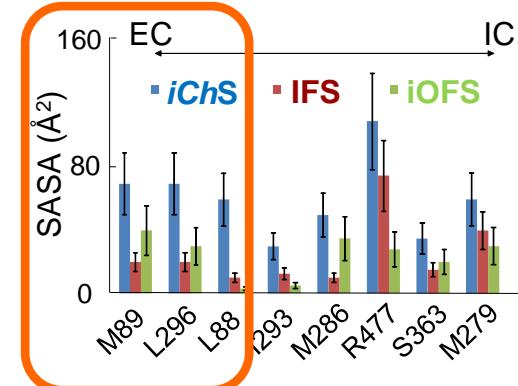
open-channel



solvent accessibility
 Glt_{Ph}



hEAAT1

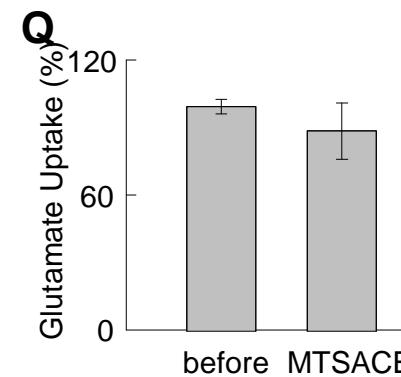
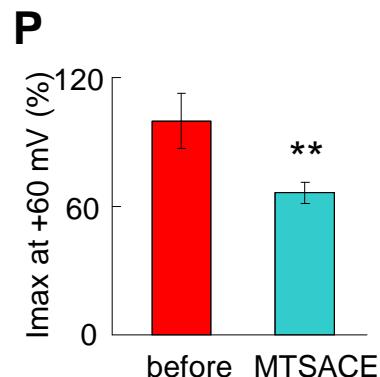
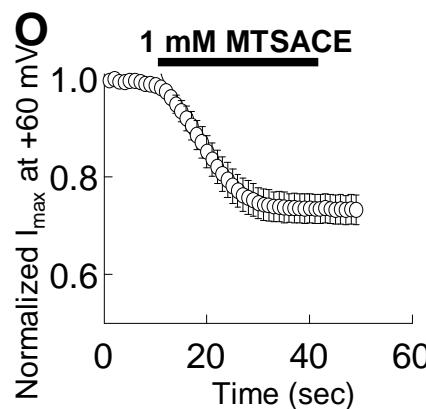
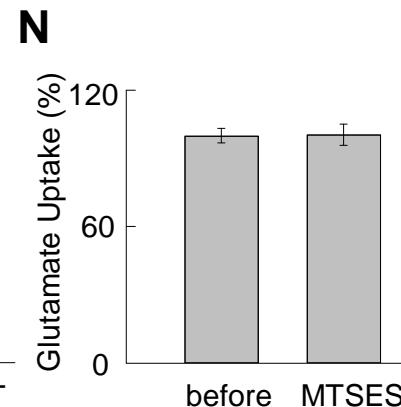
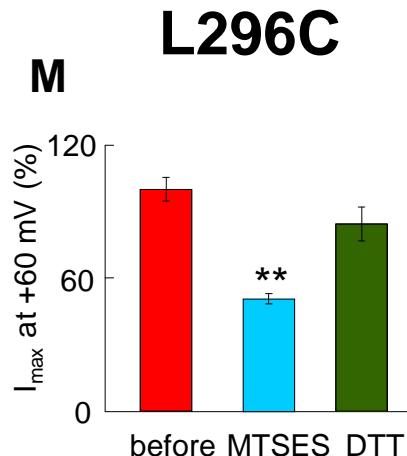
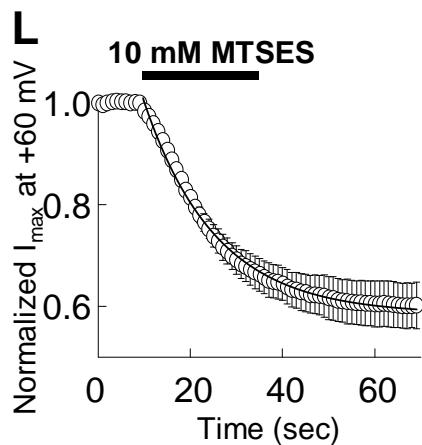


Experiments confirm the key role of L296

SCAM

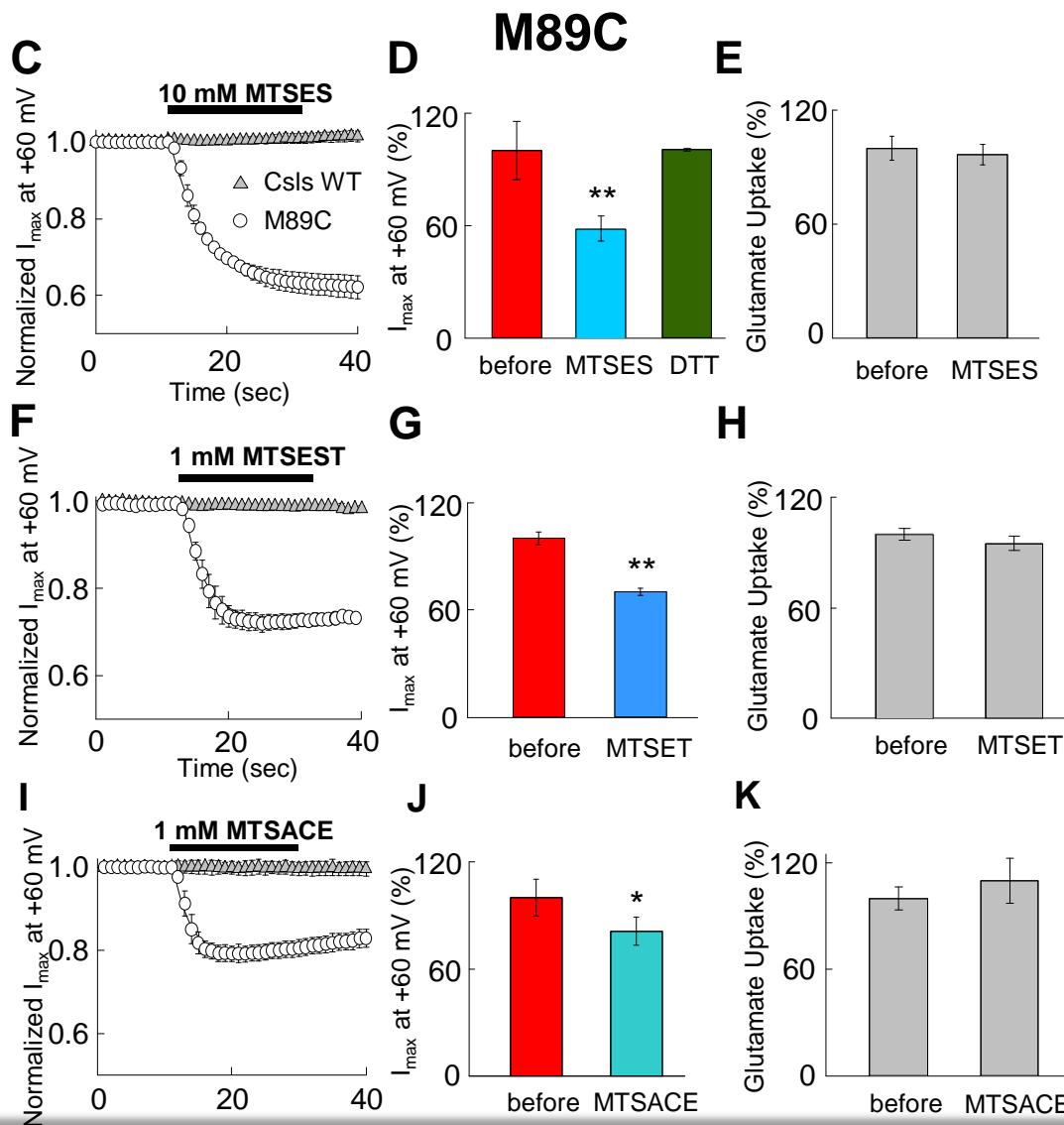
Electrophysiology

Glu uptake



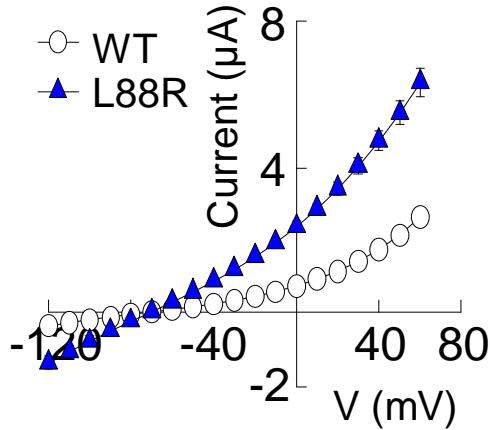
MTS-reagents that are covalently bound to cysteines: MTSES :negatively-charged; MTSET: positively-charged MTSAE : neutral DTT: reagent to reduce the disulfide bridge

M89 in EAAT1 also controls anion permeation

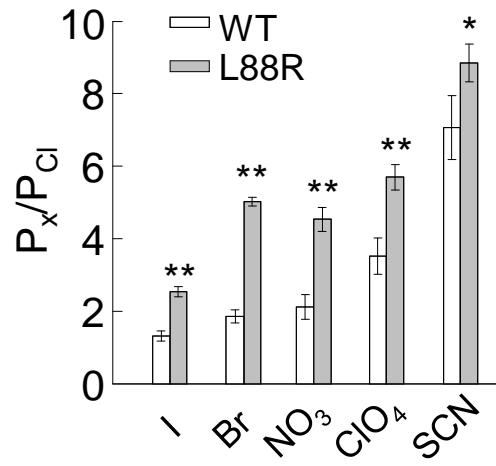


L88 controls both anion permeation and substrate transport

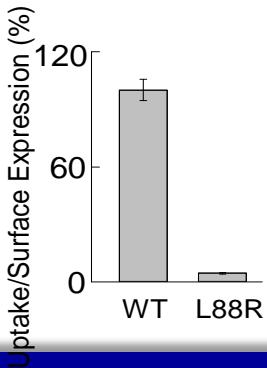
Current-voltage



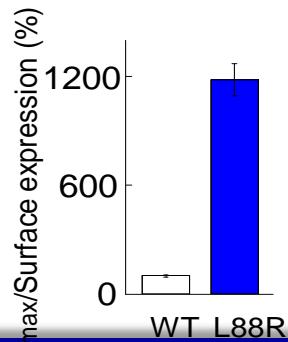
ion selectivity



Glu uptake



Normalized I_{max}



Conclusion

- Anion permeation takes place in the intermediate channeling state, *iChS*.
- Channel opening is enabled by
 - elevator-like downward movement of transport core in the substrate-loaded state
 - repacking of a cluster of hydrophobic residues (L88, M89 and L296 in EAAT1)
- Robustly shared by both the archaeal and mammalian transporters.